



Genome Assembly

李余动

lyd@zjsu.edu.cn

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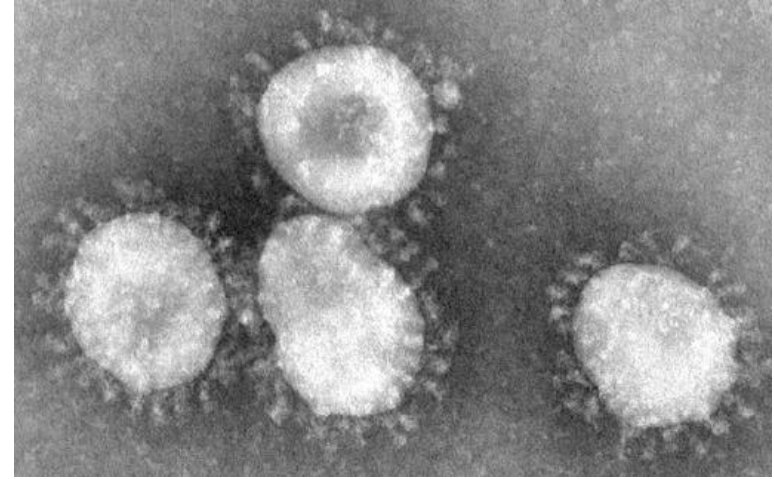
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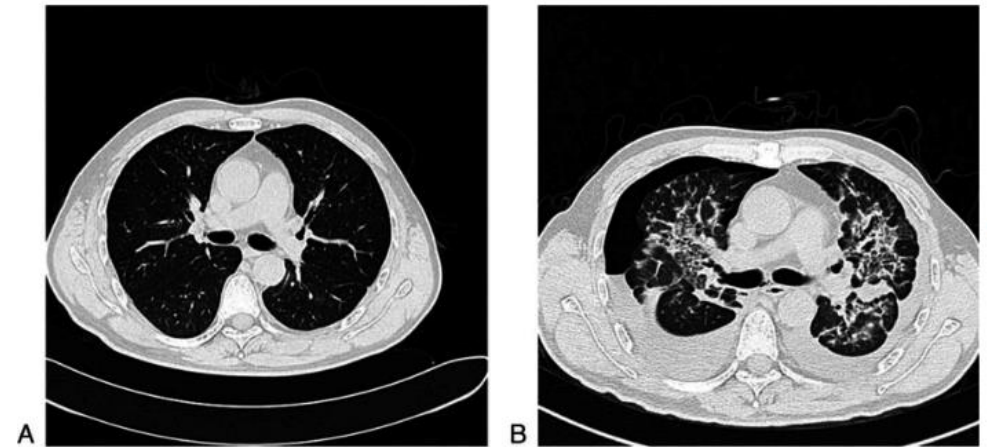
新冠病毒概述

SARS and COVID-19

- 肺炎(pneumonia):
 - Severe Acute Respiratory Syndrome (SARS), 2003
 - Coronavirus disease 2019 (COVID-19)
- SARS vs. COVID-19
 - SARS was more lethal than COVID-19, 10% of people infected with SARS died.
 - There were no **asymptomatic** SARS patients. however, COVID-19 patients sometimes without symptoms, and spread faster.



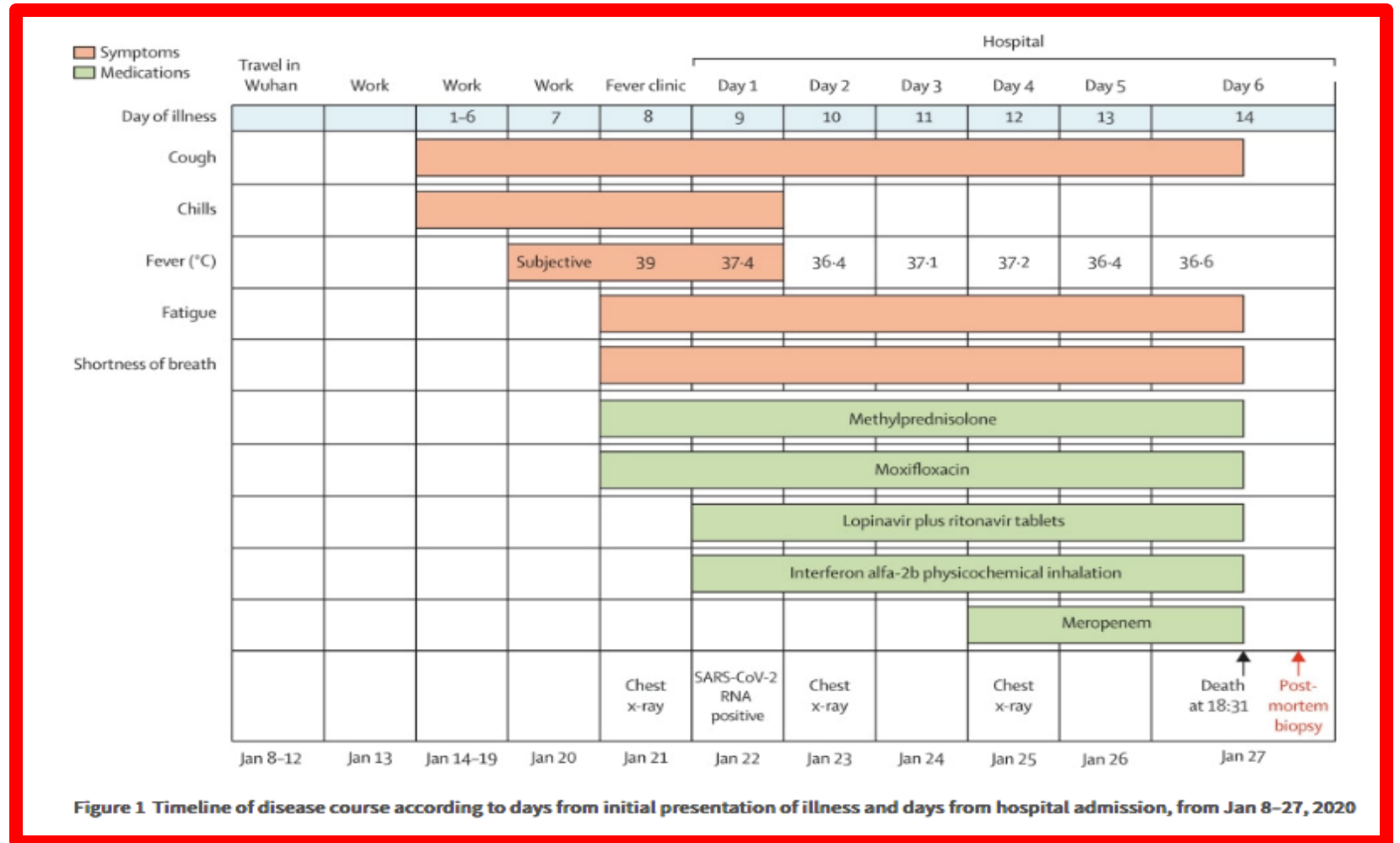
Coronavirus particles



A. Normal lung B. Lung with ground glass opacity

A COVID-19 Patient

- A 50-year-old man was admitted to a clinic in Hong Kong on Jan 21, 2020.
- Symptoms of fever, chills, cough, fatigue and shortness of breath.
- He traveled to Wuhan Jan 8–12.
- Initial symptoms of mild chills and dry cough on Jan 14 (day 1 of illness).
- Did not see a doctor and kept working until January 21.
- Chest x-ray showed multiple patchy shadows in both lungs.
- On January 22 (day 9 of illness), confirmed by reverse real-time PCR assay that the patient had COVID-19.



Article

A new coronavirus associated with human respiratory disease in China

<https://doi.org/10.1038/s41586-020-2008-3> Fan Wu^{1,7}, Su Zhao^{2,7}, Bin Yu^{3,7}, Yan-Mei Chen^{1,7}, Wen Wang^{4,7}, Zhi-G Zhao-Wu Tao², Jun-Hua Tian³, Yuan-Yuan Pei¹, Ming-Li Yuan², Yu-Li Yi Liu¹, Qi-Min Wang¹, Jiao-Jiao Zheng¹, Lin Xu¹, Edward C. Holmes

Received: 7 January 2020

Article

A pneumonia outbreak associated with a new coronavirus of probable bat origin

<https://doi.org/10.1038/s41586-020-2012-7>

Received: 20 January 2020

Accepted: 29 January 2020

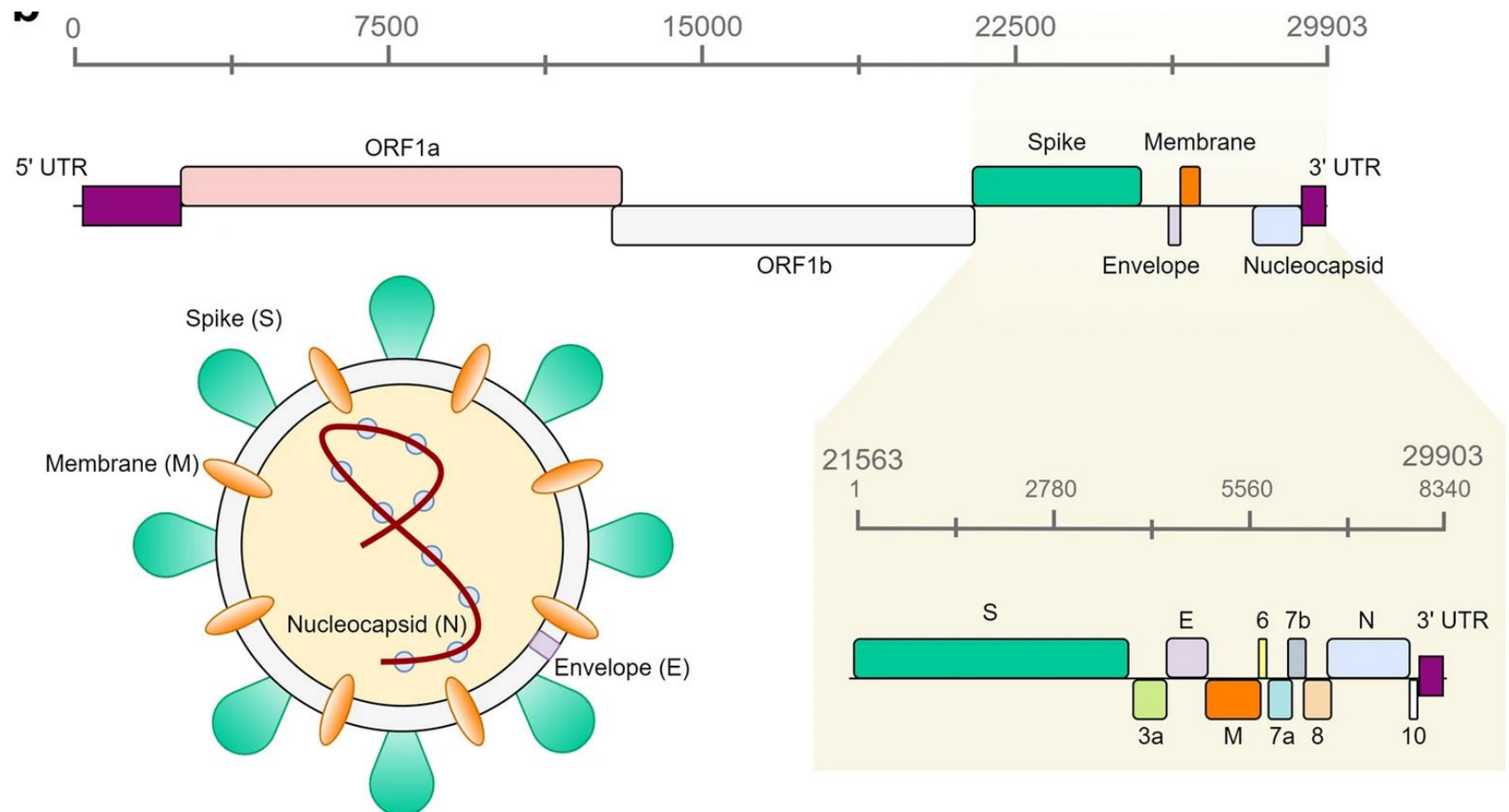
Published online: 3 February 2020

Peng Zhou^{1,5}, Xing-Lou Yang^{1,5}, Xian-Guang Wang^{2,5}, Ben Hu¹, Lei Zhang¹, Wei Zhang¹, Hao-Rui Si^{1,3}, Yan Zhu¹, Bei Li¹, Chao-Lin Huang², Hui-Dong Chen², Jing Chen^{1,3}, Yun Luo^{1,3}, Hua Guo^{1,3}, Ren-Di Jiang^{1,3}, Mei-Qin Liu^{1,3}, Ying Chen^{1,3}, Xu-Rui Shen^{1,3}, Xi Wang^{1,3}, Xiao-Shuang Zheng^{1,3}, Kai Zhao^{1,3}, Quan-Jiao Chen¹, Fei Deng¹, Lin-Lin Liu⁴, Bing Yan¹, Fa-Xian Zhan⁴, Yan-Yi Wang¹, Geng-Fu Xiao¹ & Zheng-Li Shi^{1,5}

- 2020年初复旦大学与武汉病毒所研究团队分别对患者支气管肺泡灌洗液进行了mNGS测序，鉴定出了一株新型冠状病毒（Coronavirus）。
- 根据病毒基因组的系统发育分析发现，该病毒基因组与蝙蝠体内发现的类严重急性呼吸综合征（SARS-like）冠状病毒基因组的相似性高达89.1%。
- 新冠病毒的全基因组序列对新冠疫情的防疫工作具有重大意义，如设计测序引物开展核酸检测试剂盒开发，或根据新冠刺穿蛋白的基因序列进行疫苗研发等。

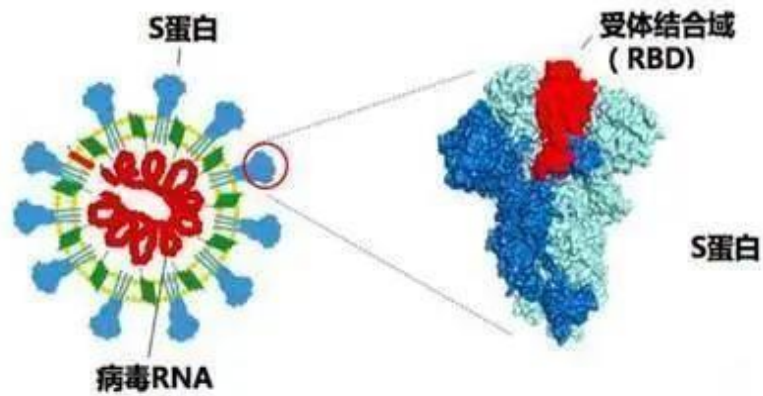
The Coronavirus genome

- RNA virus (single-stranded, positive-sense)
- Linear genome = ~30,000 nucleotides
- 11 coding-regions (genes)
- 12 potential gene productions

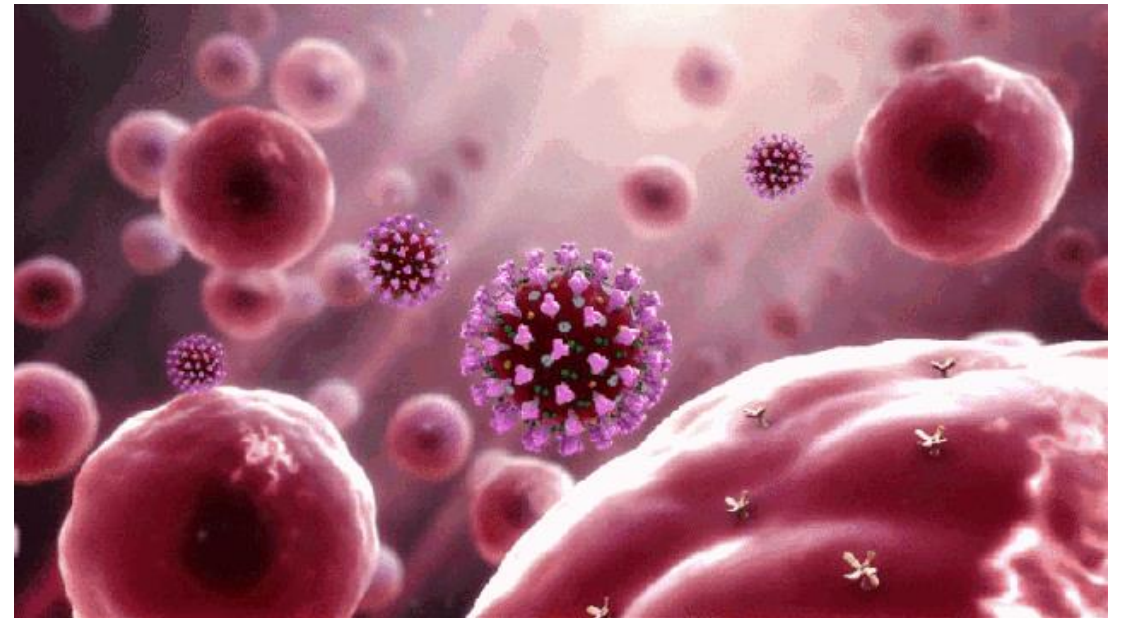


A CLOSE UP OF THE VIRUS

新冠病毒是一种RNA病毒
外壳上是刺突状的S蛋白，其头部是受体结合域 (RBD)

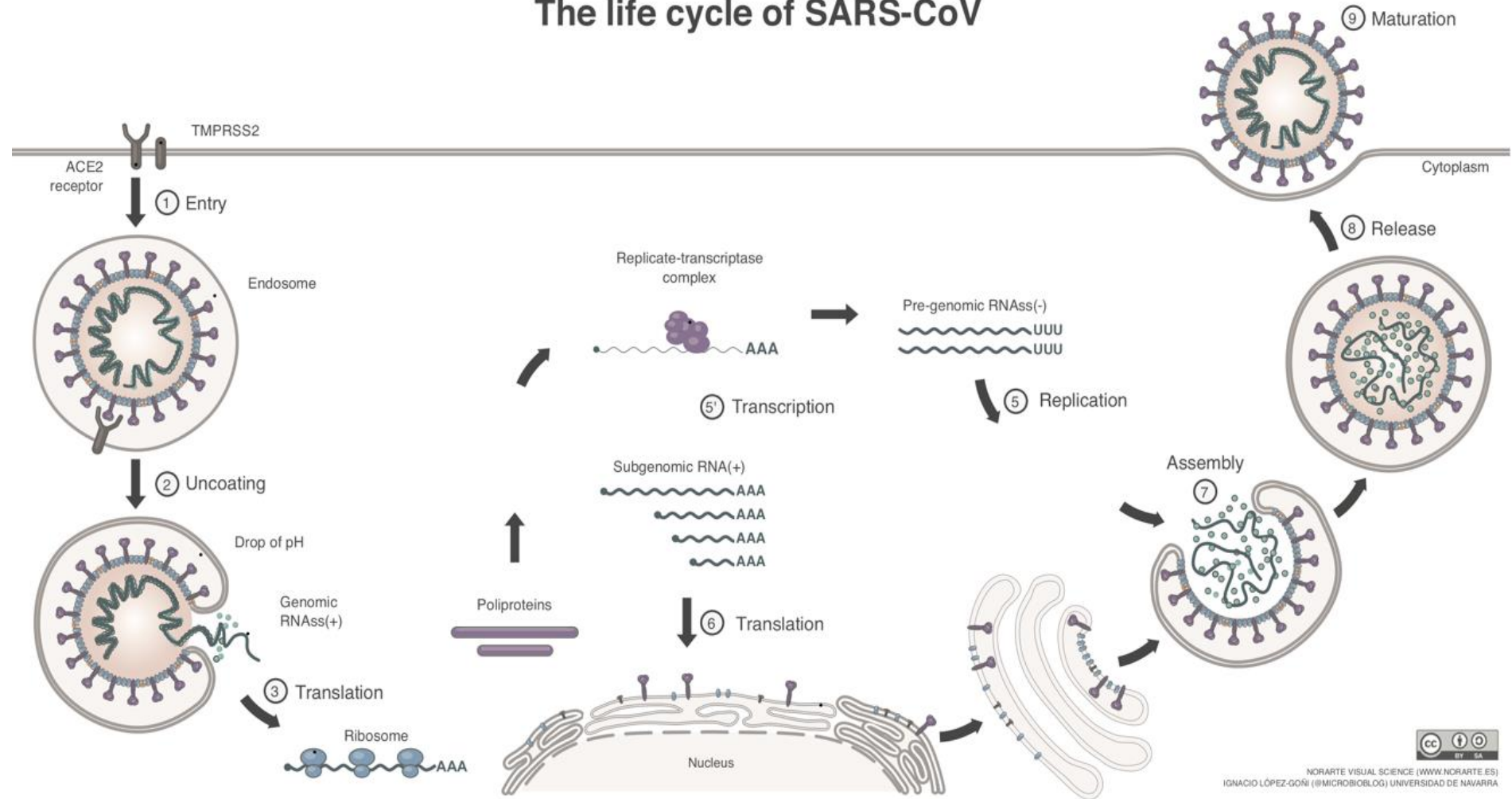


Viral spikes target angiotensin converting enzyme 2 (ACE2) receptors—found in lung pneumocytes II (for one).



The Life cycle of SARS-CoV-2

The life cycle of SARS-CoV



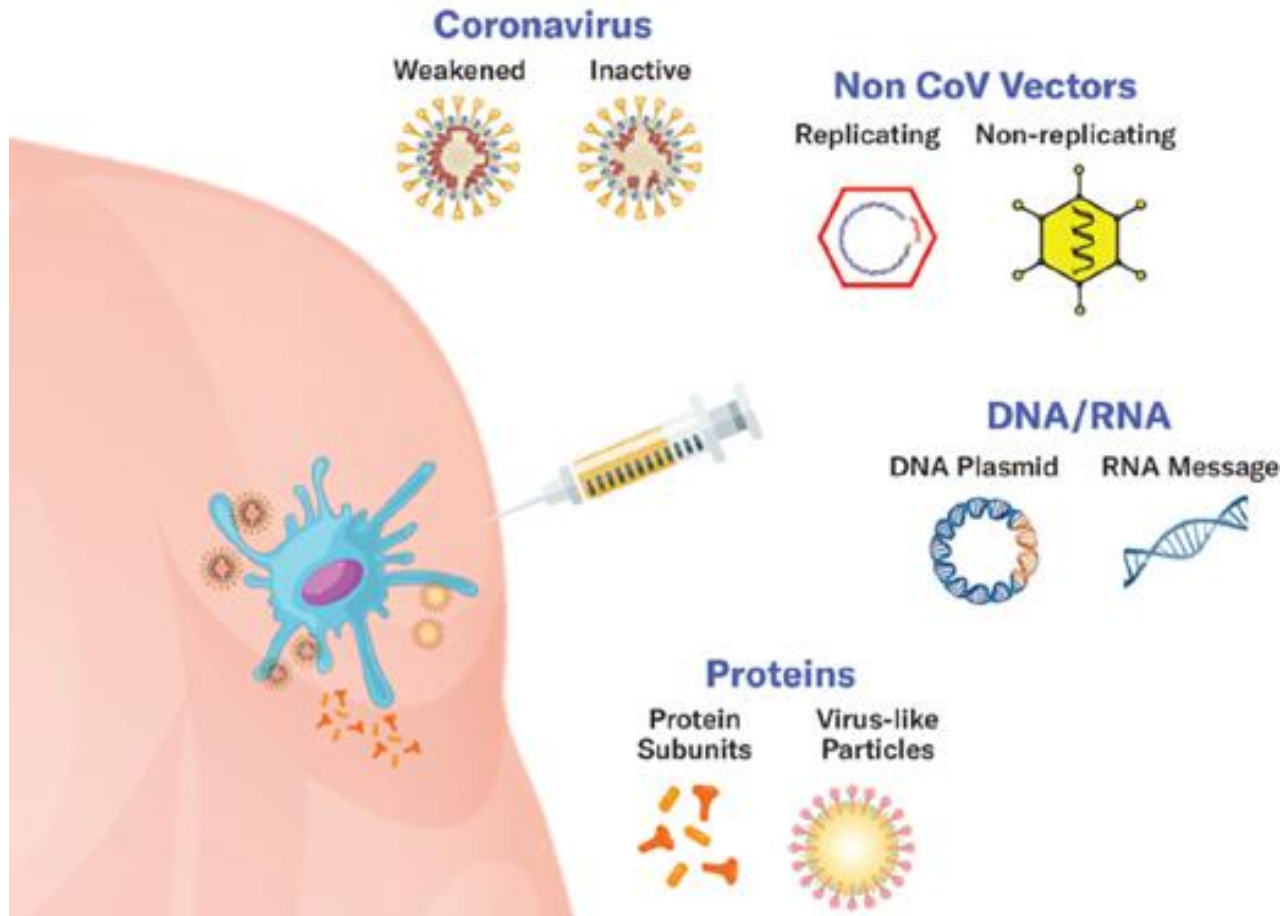
- Polyprotein
- Protease

https://upload.wikimedia.org/wikipedia/commons/c/ca/SARS-CoV-2_ciclo.svg

Virus proteins and vaccine

疫苗是病原体的一部分蛋白质(也叫抗原), 可刺激机体产生抗体。

SARS-CoV-2 Vaccine Approaches

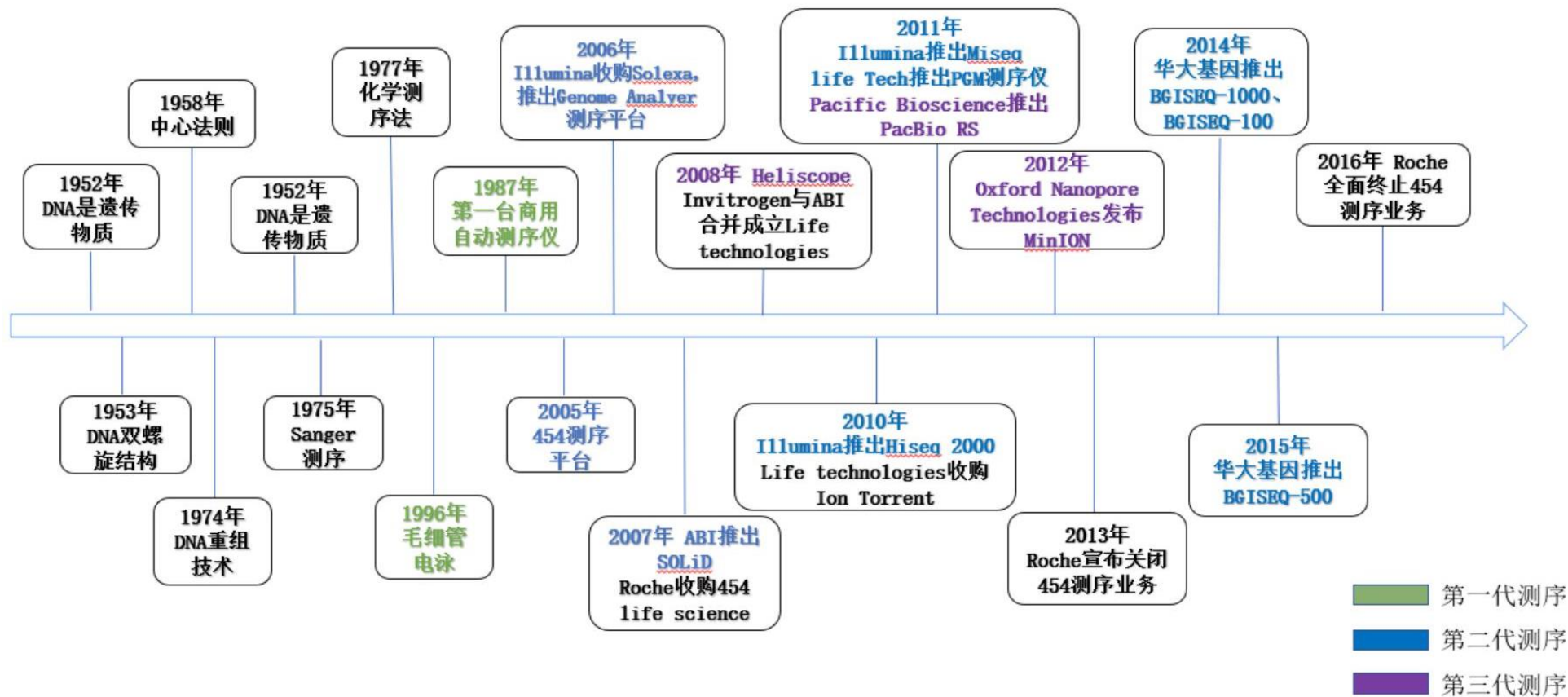


- 灭活疫苗
- 减毒疫苗
- 亚单位疫苗
- 核酸疫苗
 - RNA vaccine (Pfizer, Moderna)



基因组测序策略

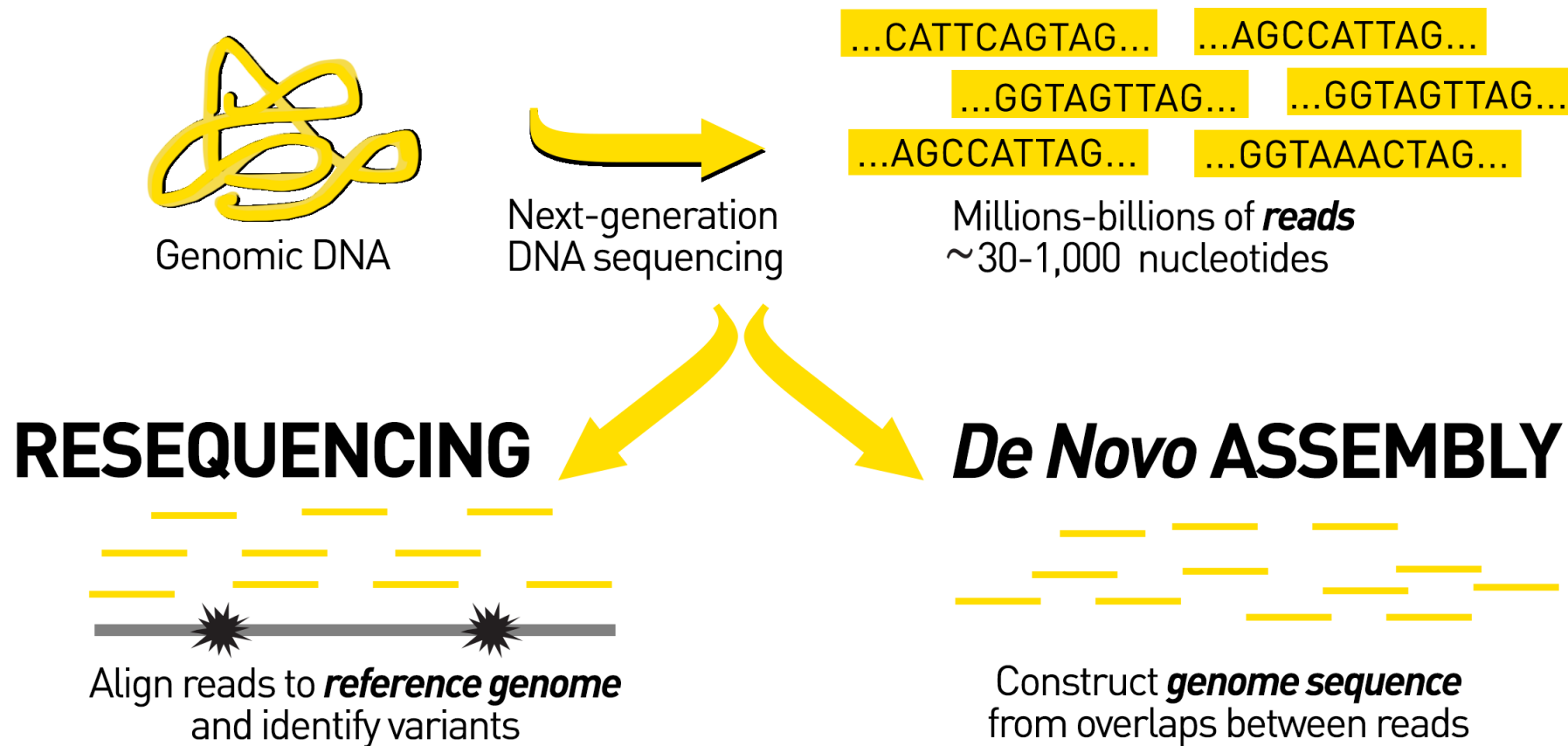
DNA测序技术发展历程



	一代	二代	三代
并行度	低<400	高>10000	
PCR	需要		不需要

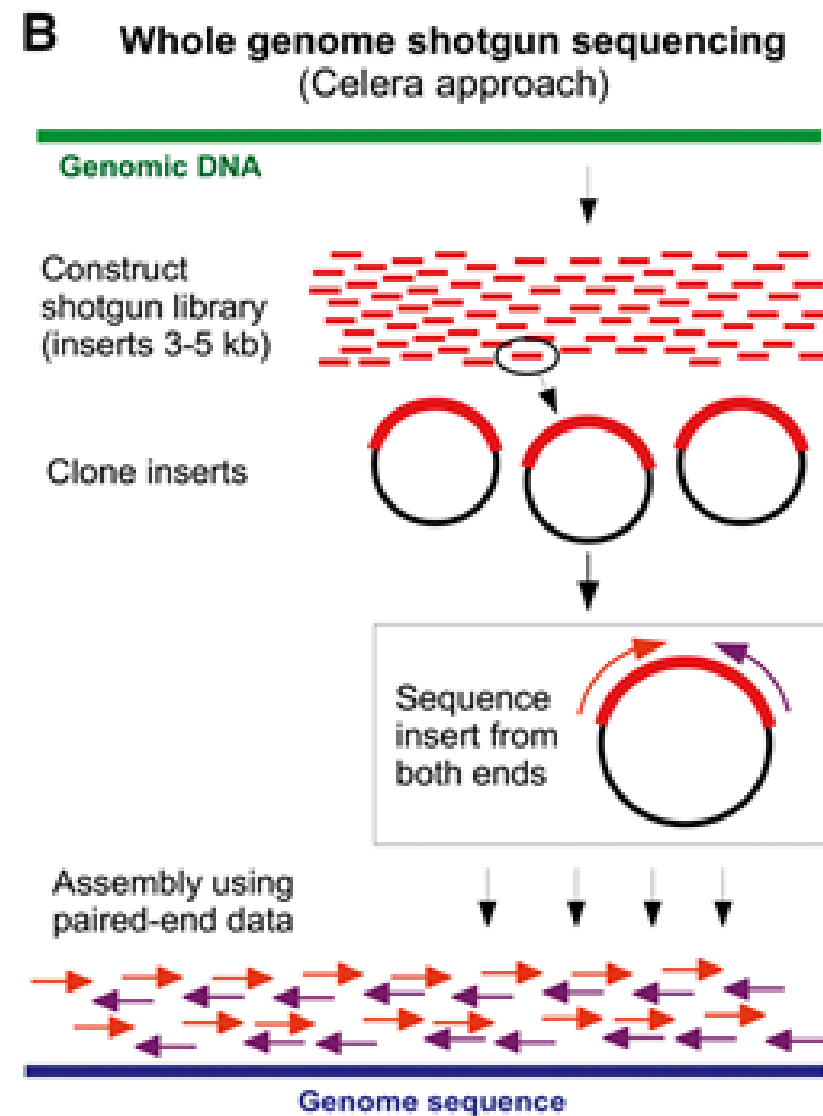
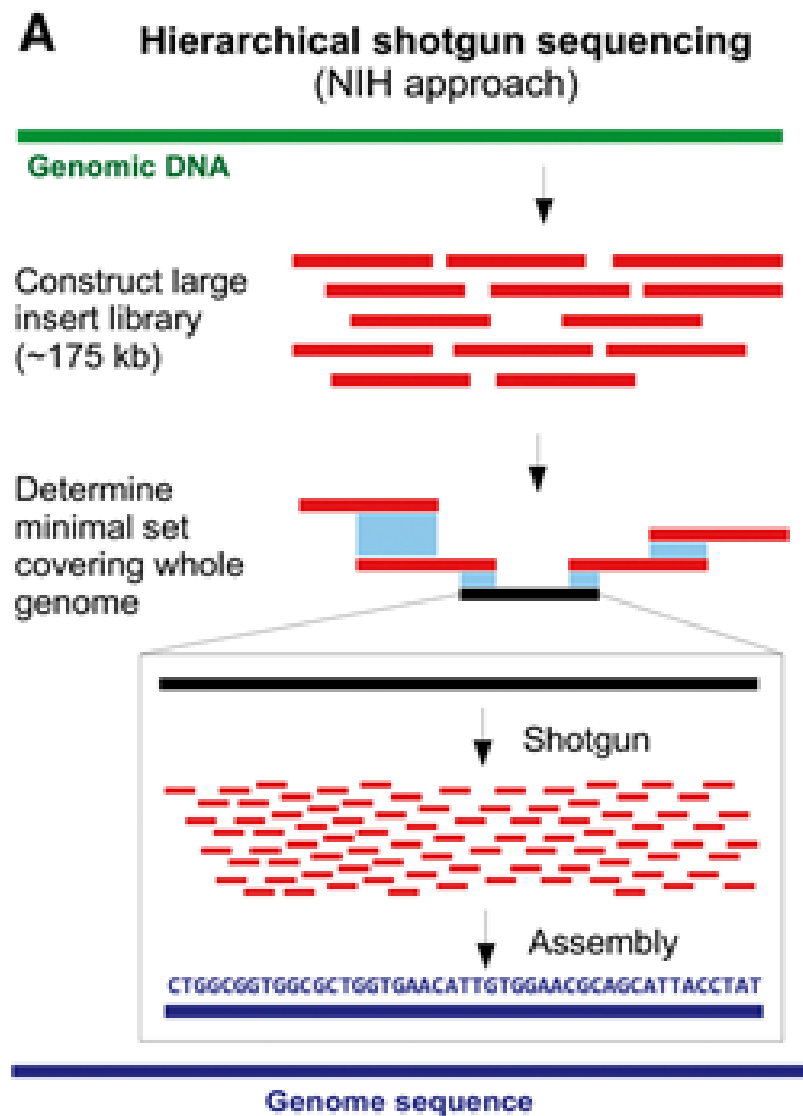
全基因组测序(Whole Genome Sequencing)

- 重测序(RESEQUENCING): 将测序reads比对到参考基因组, 并鉴定遗传差异。
- 从头组装(*De Novo* Assembly): 将测序reads准确地拼接成基因组。



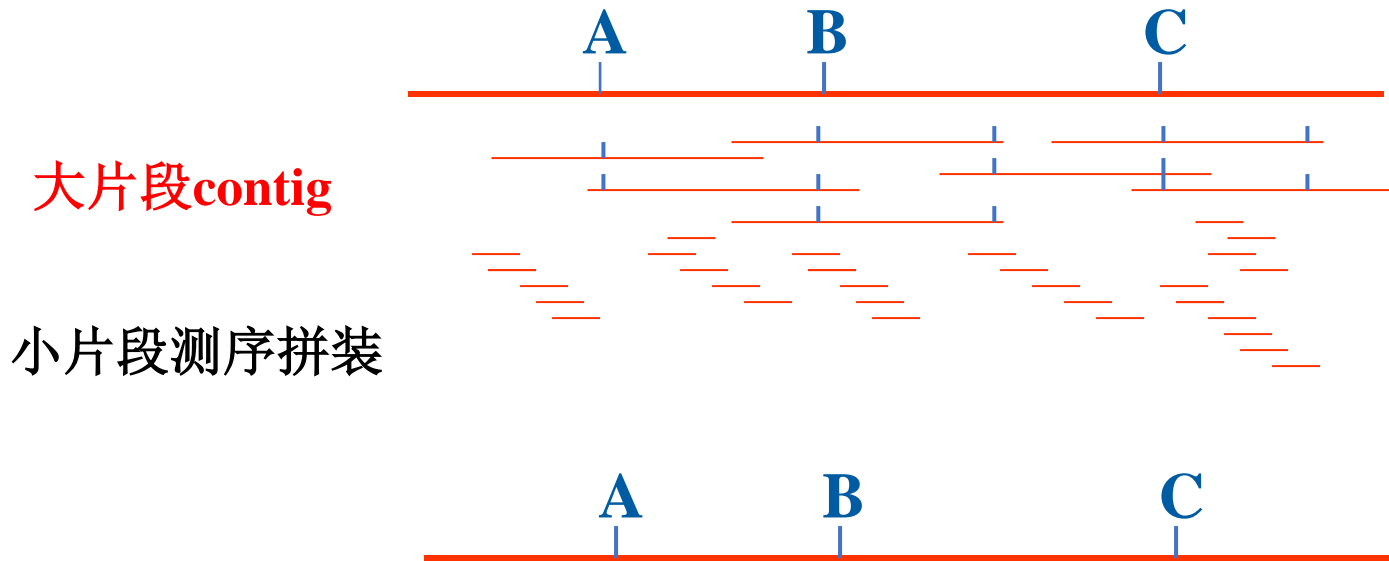
基因组测序两种策略

- 逐步克隆法(A)
- 全基因组鸟枪法(B)



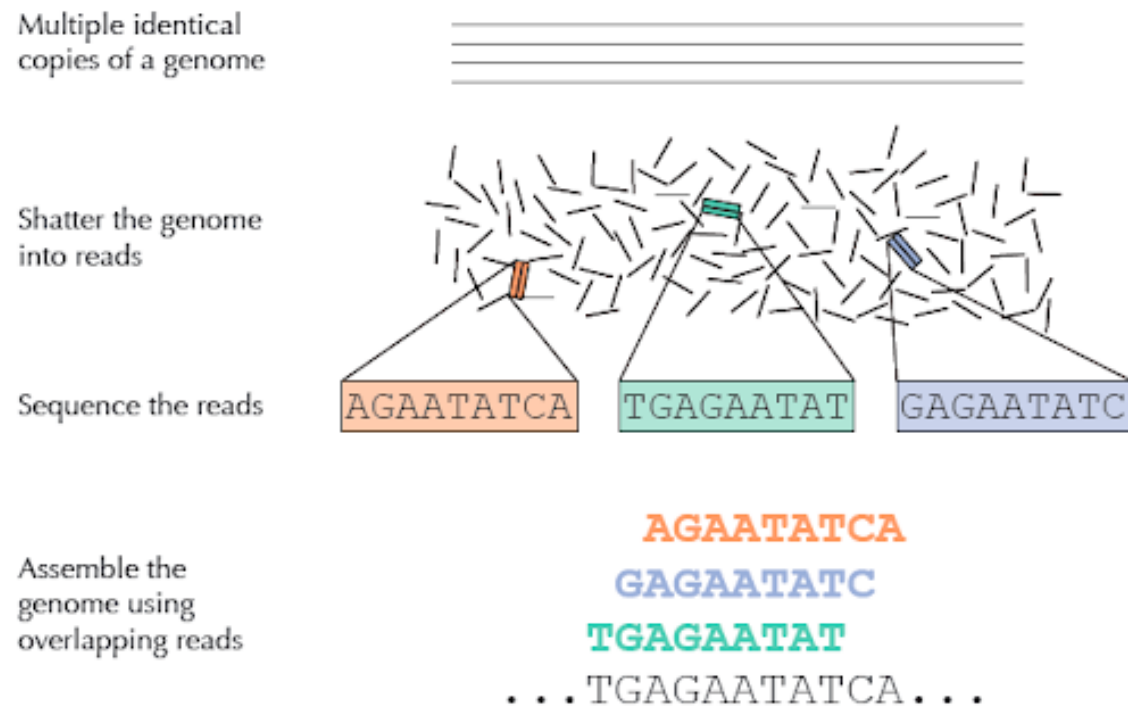
基于克隆群(contig-based)的策略

先将染色体打成比较大的片段(几十-几百Kb), 利用分子标记将这些大片段排成重叠的克隆群(Contig), 分别测序后拼装。

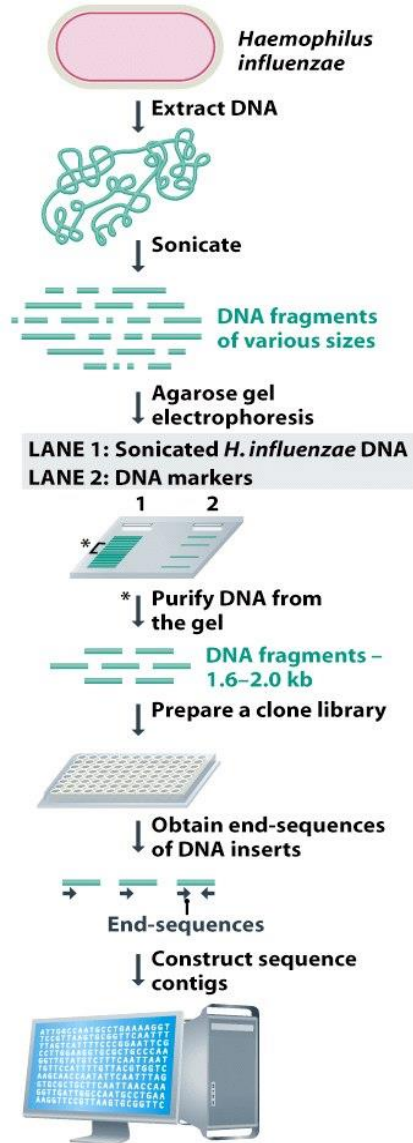


Shotgun测序策略—序列的组装

- 序列组装原理: 直接从已测序的小片段中寻找彼此重叠的测序克隆, 然后依次向两侧邻接的序列延伸。



实例: 流感嗜血杆菌基因组的测序及序列组装



超声波打断纯化的基因组DNA

琼脂糖电泳收集1.6~2.0Kb的区段、纯化

构建到质粒载体中

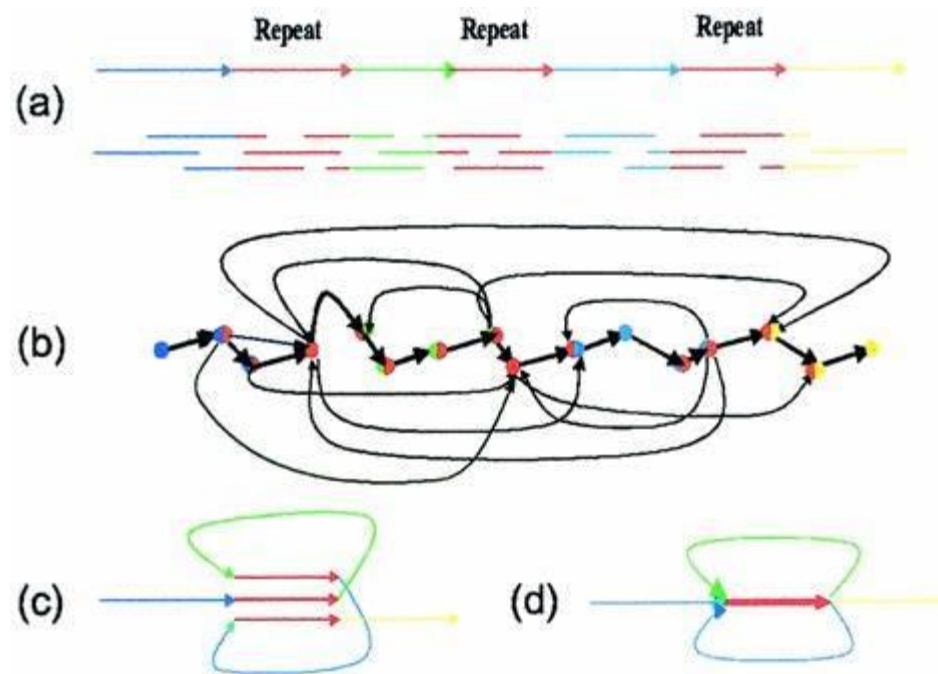
随机挑选19687个克隆,进行28643次测序,得到可读顺序为11 631 485 bp

组装成140个覆盖全基因组范围的独立的顺序重叠群,

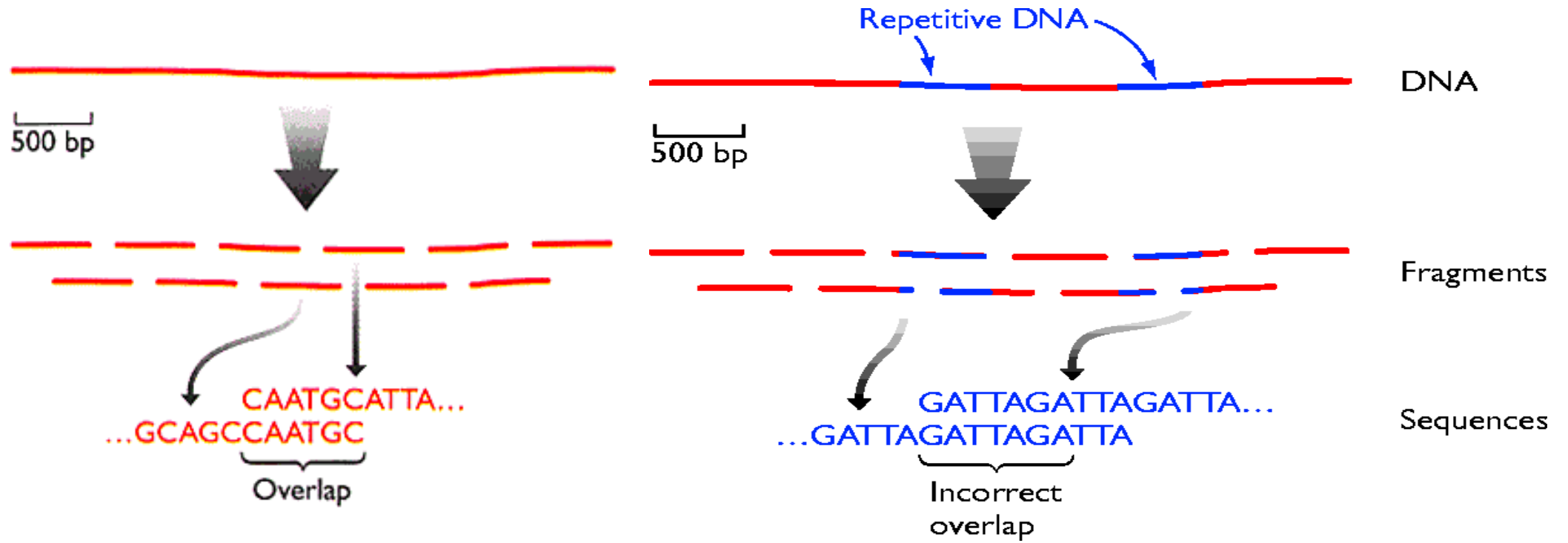
.....完整基因组

Shotgun测序—拼接问题？

- 因为整个基因组太长 ($>1M$), 而每次只能测得一个500的小片断(read)
- 问题: 如何根据read恢复原始顺序?
- 类比: 10本圣经, 都从随机点起始剪成500个字母左右的小纸条, 问: 给你这么一堆小纸条, 你能读出圣经来吗?
- 转成图论问题: Hamilton和Euler路径
- 但是都会拼错!



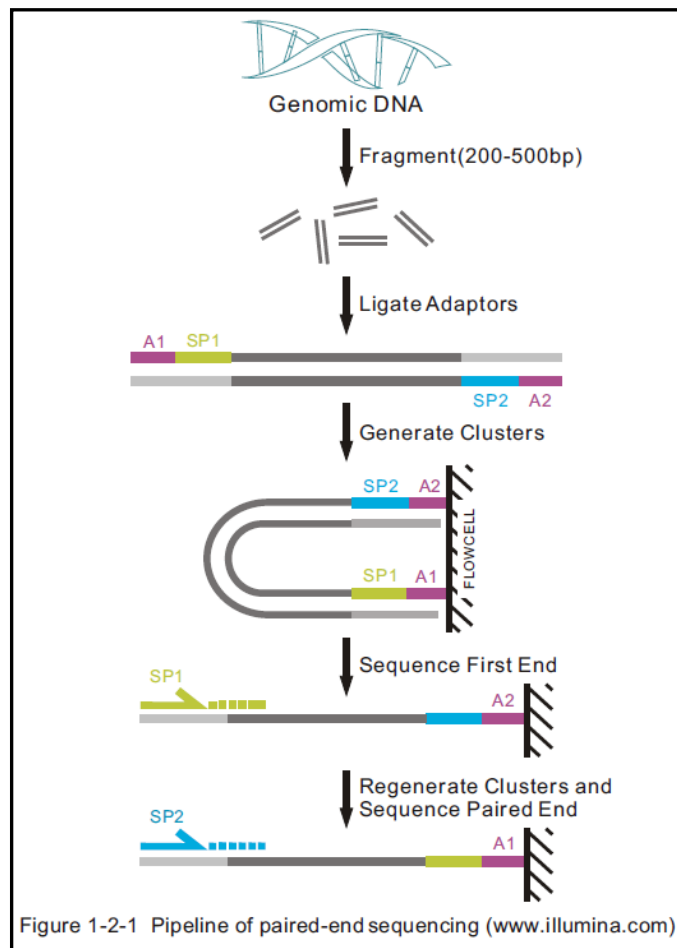
拼接错误：Repeat的存在



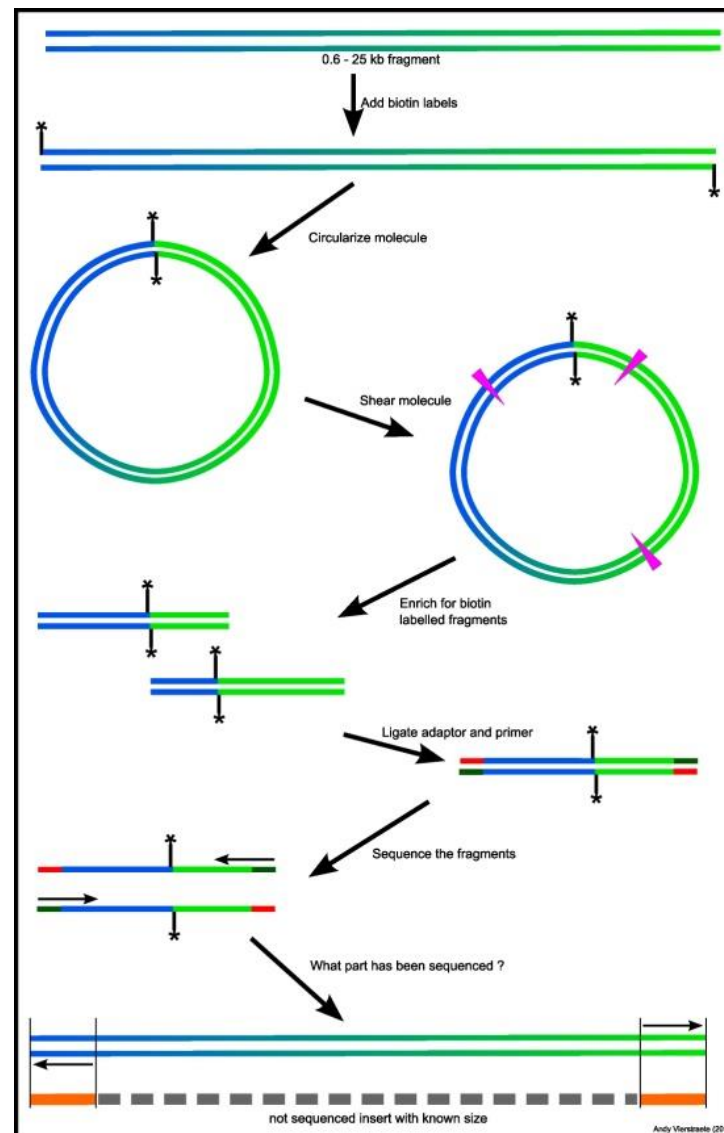
基因组重复序列测序策略

二代双端测序可通过构建长文库，跨过一个大片段序列的两末端。

双端测序Paired-End Sequencing

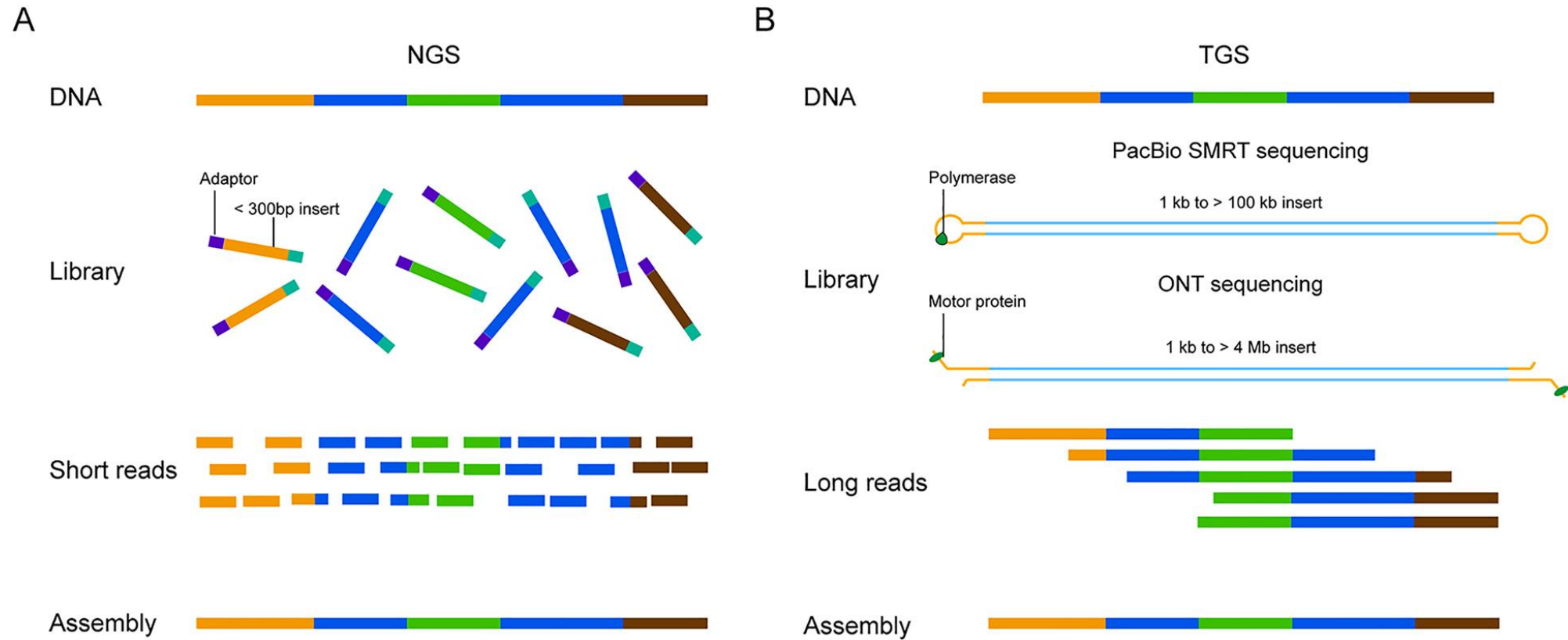


环化测序Mate-Pair Sequencing



基因组重复序列测序策略

随着三代测序技术的进步，基因组测序一般采用二代三代组合测序策略



Chen Z, He X. Application of third-generation sequencing in cancer research. Medical Review. 2021 Dec 1;1(2):150–71.



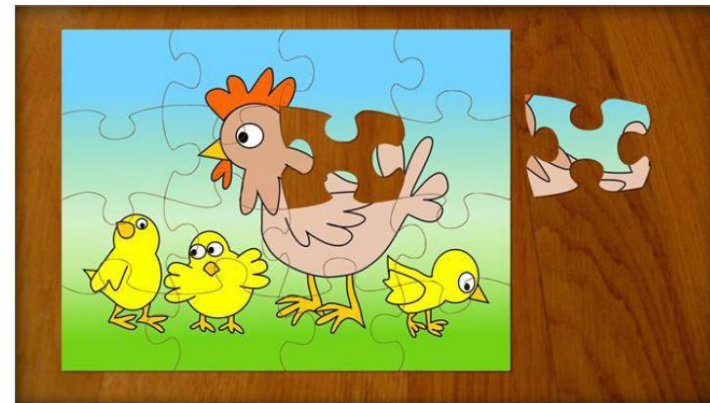
Genome Assembly

De novo Assembly

- 从头组装：把短序列组装起来，拼出一条完整的测序序列
- 从头组装类似拼图
 - The bigger the pieces the easier to reconstruct the puzzle!

```
>NC_045512.2 Severe acute respiratory syndrome coronavirus 2 isolate Wuh  
complete genome  
ATTAAAGGTTTATACCTTCCCAGGTAACAAACCAACCAACTTTCGATCTCTTGATAGATCTGTTCTCTAAA  
CGAACTTTAAAATCTGTGTGGCTGTCACTCGGCTGCATGCTTAGTGCACTCACGCAGTATAATTAATAAC  
TAATTACTGTCGTTGACAGGACACGAGTAACCTGCTATCTTCTGCAGGCTGCTTACGGTTTCGTCCGTG  
TTGCAGCCGATCATCAGCACATCTAGGTTTCGTCCGGGTGTGACCGAAAGGTAAGATGGAGAGCCTTGTC  
CCTGGTTTCAACGAGAAAACACACGTCCAACCTCAGTTTGCCGTGTTTTACAGGTTTCGCGACGTGCTCGTAC  
GTGGCTTTGGAGACTCCGTGGAGGAGGCTTATCAGAGGCACGTCAACATCTTAAAGATGGCACTTGTGG  
CTTAGTAGAAGTTGAAAAAGGCGTTTTGCCTCAACTTGAACAGCCCTATGTGTTTCAAAACGTTTCGGAT  
GCTCGAAGTGCACCTCATGGTCATGTTATGGTTGAGCTGGTAGCAGAAGTGAAGGCATTTCAGTACGGTC  
GTAGTGGTGAGACACTTGGTGTCTTGTCCCTCATGTGGGCGAAATACCAGTGGCTTACCAGCAAGGTTCT  
TCTTCGTAAGAACGGTAATAAAGGAGCTGGTGGCCATAGTTACGGCGCCGATCTAAAGTCATTGACTTA  
GGCGACGAGCTTGGCACTGATCCTTATGAAGATTTTCAAGAAAAGTGAACACTAAACATAGCAGTGGTG  
TTACCCGTGAACTCATGCGTGAGCTTAAACGGAGGGGCATACACTCGCTATGTCGATAACAACCTTCTGTGG  
CCCTGATGGTACCCTCTTGAAGTGCATTAAGACCTTCTAGCACGTGCTGGTAAAGCTTTCATGCACTTTG  
TCCGAACAAGTGGACTTTATTGACACTAAGAGGGGTGTATACTGCTGCCGTGAACATGAGCATGAAATTG  
CTTGGTACACGGAACGTTCTGAAAAGAGCTATGAATTGCAGACACCTTTTAAAATTAATTGGCAAAGAA  
ATTTGACACCTTCAATGGGGAATGTCCAAATTTGTATTTCCCTTAAATTCATAATCAAGACTATTCAA  
CCAAGGGTTGAAAAGAAAAGCTTGATGGCTTATGGGTAGAATTCGATCTGTCTATCCAGTTGCGTCAC
```

.fasta consensus genome



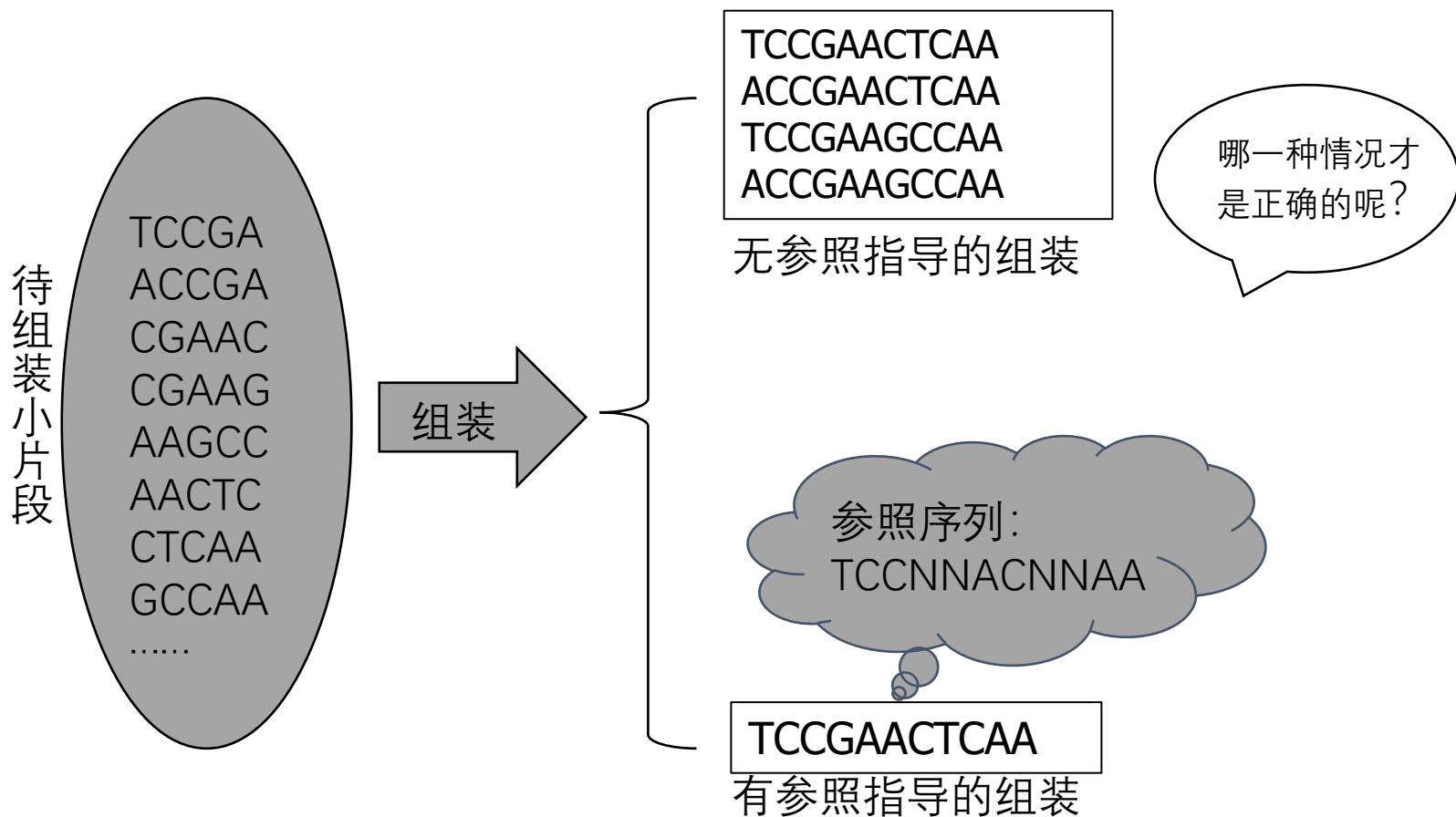
基因组组装方式

- *De novo* assembly: without reference sequences

- 测序一个新的物种的基因组

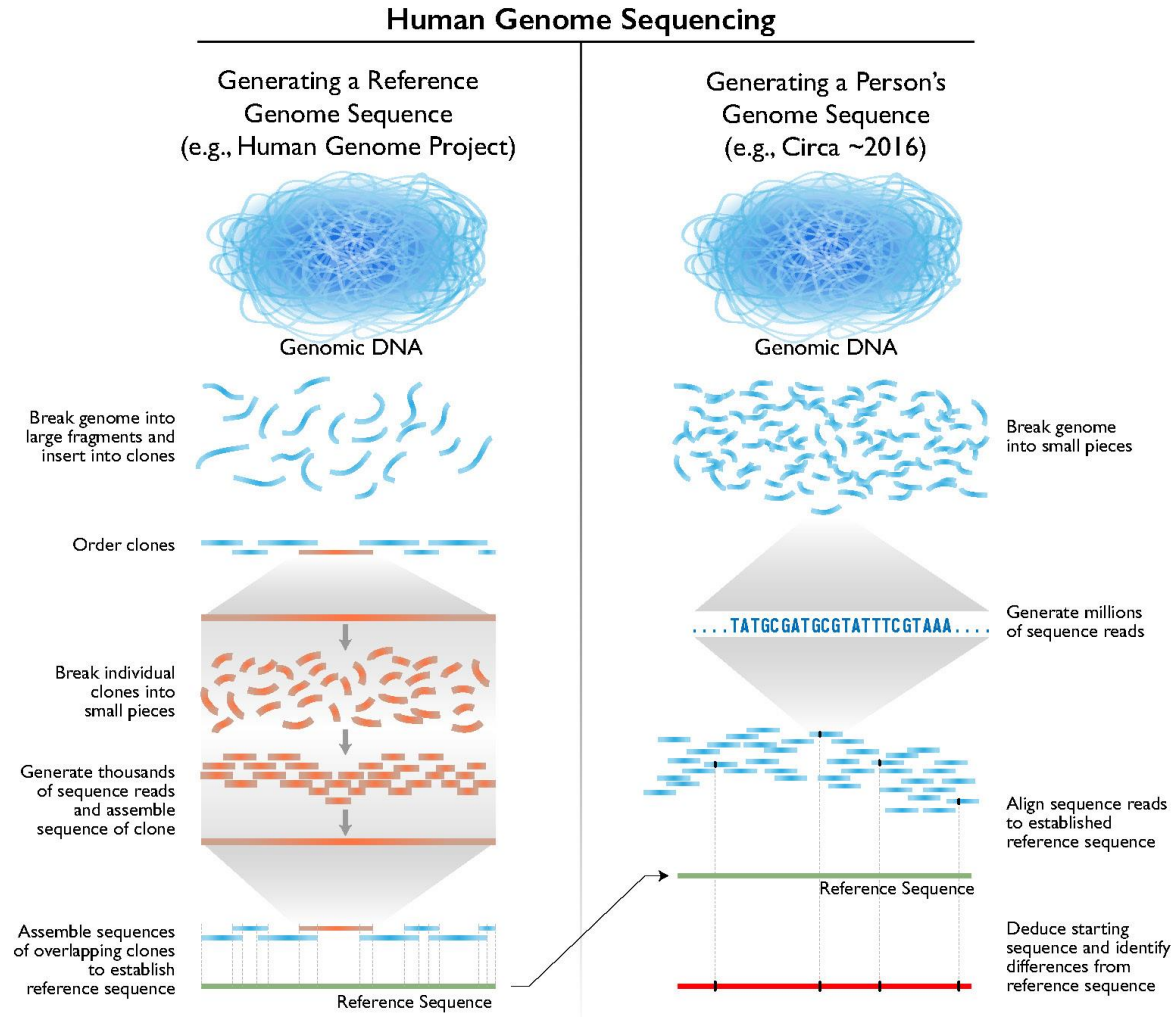
- Reference-guided assembly: with reference sequences

- 测序相近的物种（亚种）的基因组
- 测序数据mapping到参考序列上，推测可能的基因组



参照序列用于指导序列片段组装

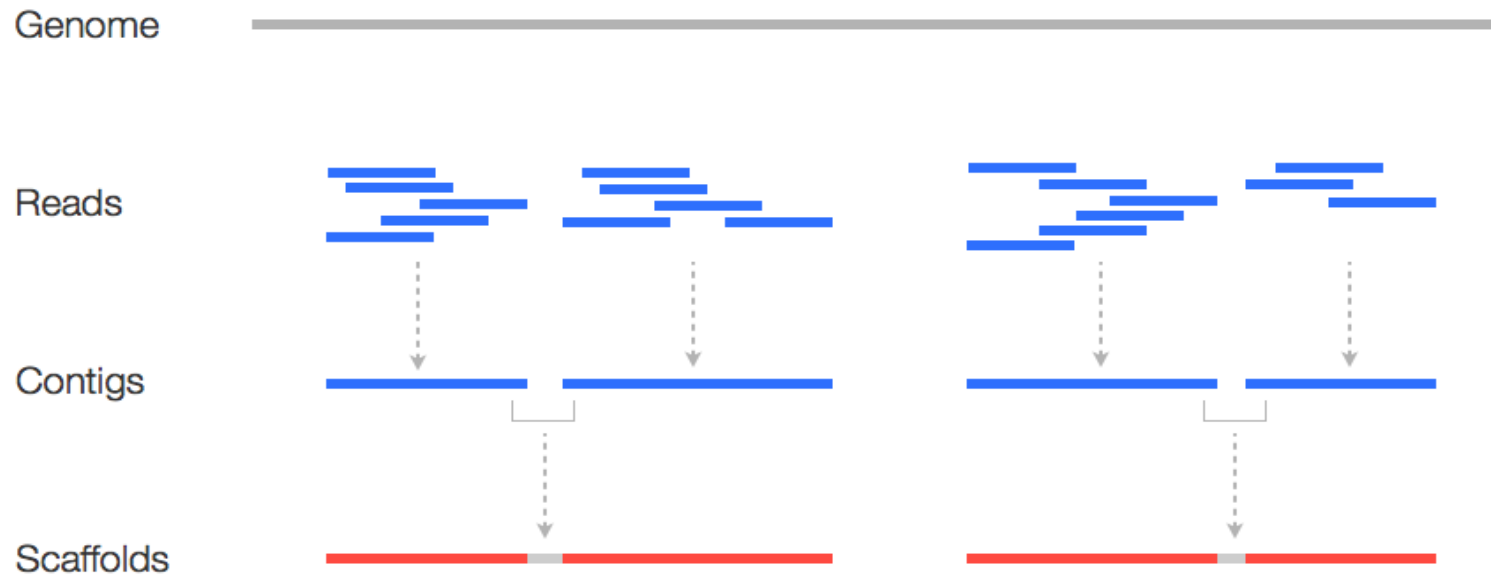
人类基因组测序方法



个人基因组测序

<https://www.genome.gov/about-genomics/fact-sheets/Sequencing-Human-Genome-cost>

De novo assembly



基因组组装的概念:

- Reads: 读段，即测序产生的短序列，通常一代的reads读长在一千左右，二代的reads相对较短，平均是几百bp，三代的reads较长在几千到几万bp之间；
- Contig: 重叠群，基于reads之间的overlap区域拼接成的连续序列。
- Scaffold: 获得contig按照一定顺序和方向组成scaffold，不同contig之间还有空缺(Gap)。

De novo Assembly: 工作草图与完成图/精细图

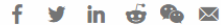
- Gap filling and assembly polishing
- Draft genome (草图): 90%
- Finished genome (精细图): 99.99%

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HOME > SCIENCE > VOL. 376, NO. 6588 > THE COMPLETE SEQUENCE OF A HUMAN GENOME

SPECIAL ISSUE RESEARCH ARTICLE | HUMAN GENOMICS



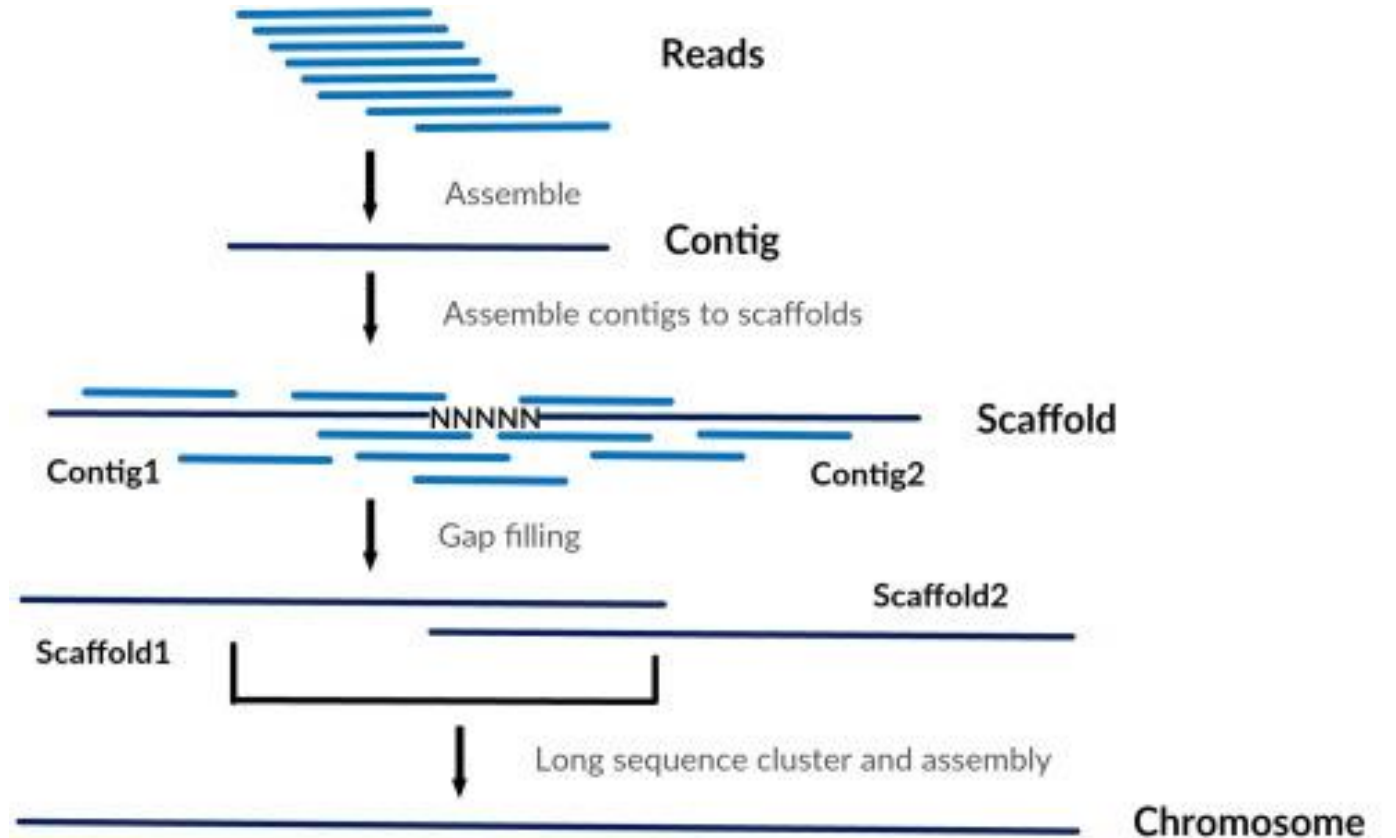
The complete sequence of a human genome

SERGEY NURK, SERGEY KOREN, ARANG RHIE, MIKKO RAUTIAINEN, ANDREY V. BZIKADZE, ALLA MIKHEENKO,

MITCHELL R. VOLLGER, NICOLAS ALTEMOSE, LEV URALSKY, (...) ADAM M. PHILLIPPY, +91 authors

[Authors Info & Affiliations](#)

SCIENCE • 31 Mar 2022 • Vol 376, Issue 6588 • pp. 44-53 • DOI: 10.1126/science.abbj6987



De novo Assembly的两种常用算法

● Overlap-Layout-Consensus (OLC) – 序列重叠一致性

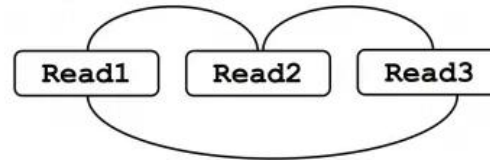
- Phrap
- Newbler
- Canu

● de Bruijn graph(DBG) – 德布鲁因图

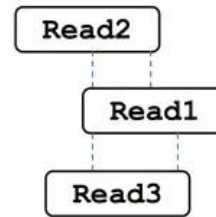
- SPAdes
- Velvet
- ABySS

(a) Overlap, Layout, Consensus assembly

(i) Find overlaps



(ii) Layout reads



(iii) Build consensus

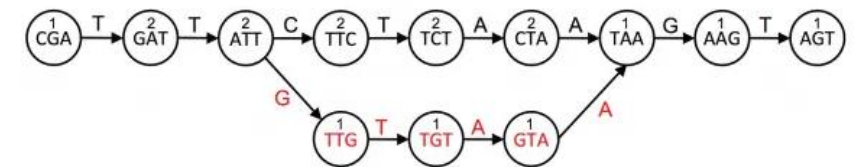
```
CGATTCTA
  TTCTAAGT
  GATTGTAA
-----
CGATTCTAAGT
```

(b) De Bruijn graph assembly

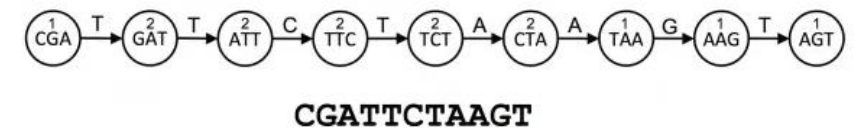
(i) Make kmers

Read1: TTCTAAGT	Read2: CGATTCTA	Read3: GATTGTAA
Kmers: TTC	Kmers: CGA	Kmers: GAT
TCT	GAT	ATT
CTA	ATT	TTG
TAA	TTC	TGT
AAG	TCT	GTA
AGT	CTA	TAA

(ii) Build graph



(iii) Walk graph and output contigs



(引自Ayling et al., Briefings in Bioinformatics, 2019)

de Bruijn图算法 – K-mer选择

Primer | Published: 08 November 2011

How to apply de Bruijn graphs to genome assembly

[Phillip E C Compeau](#), [Pavel A Pevzner](#) & [Glenn Tesler](#)

Nature Biotechnology 29, 987–991 (2011) | [Cite this article](#)

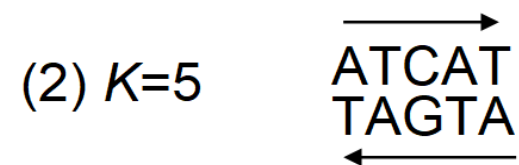
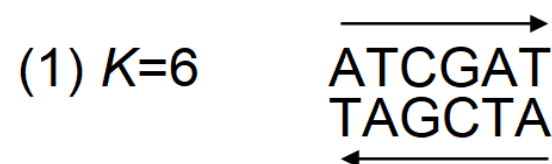
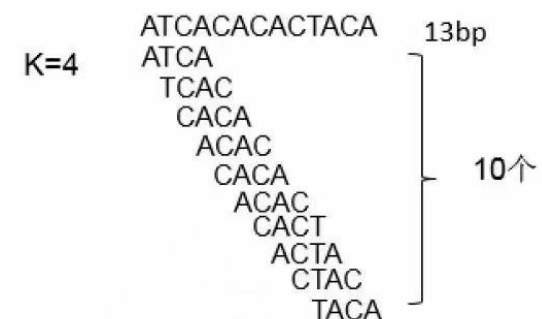
69k Accesses | 333 Citations | 107 Altmetric | [Metrics](#)

A mathematical concept known as a de Bruijn graph turns the formidable challenge of assembling a contiguous genome from billions of short sequencing reads into a tractable computational problem.

- **K-mers**指一个read中长度为k的所有子序列，如序列ATTACGTCGA可以分成一系列3-mers (k=3)：ATT, TTA, TAC, ACG, CGT, GTC, TCG和CGA。

– 若序列的长度为L，那么可以得到L-K+1个K-mers。

- 为防止由于出现回文序列而导致序列自身拼接的问题，**K值应取奇数**。



基因组组装质量评估参数

●Contig/Scaffold 数目

- 越少越好

●N50/N90

- 将Contigs/Scaffolds从长到短排序，逐项相加，长度达到全基因组一半(50%) 或90%时对应的一段Contig/Scaffold长度;
- 越长越好

●Coverage/Depth

- 测序覆盖度/深度：单个碱基被测序reads覆盖的次数

●Genome coverage

- 越长越好

测序覆盖度/深度(Coverage/Depth)

- 测序覆盖度：测序得到的总碱基数与待测基因组大小的比值。

- 测序深度：基因组上每一个位置平均覆盖读段的条数。

- 覆盖度 $C = n * l / L$

- n: 读段数

- l: 读段长度

- L: 序列长度

```
          CTAGGCCCTCAATTTTT
          CTCTAGGCCCTCAATTTTT
          GCTCTAGGCCCTCAATTTTT
TATCTCGGCTCTAGCC
          TCGGCTCTAGCCCCTCAATTT
          TCTCGGCTCTAGCCCCTC
TATCTCGGCTCTAGCCCC
          CGGTTGTAGCCCCCTAA
TATCTCGGCTCTACCC


---


TATCTCGGCTCTAGCCCCTCAATTTTT
```

9条读段
166个碱基

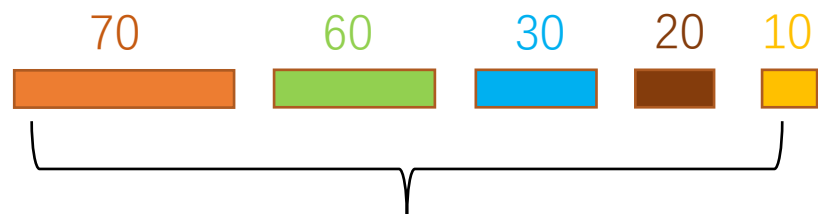
28个碱基

$$\text{覆盖度 } C = 166 / 28 = 6x$$

基因组组装质量评估参数

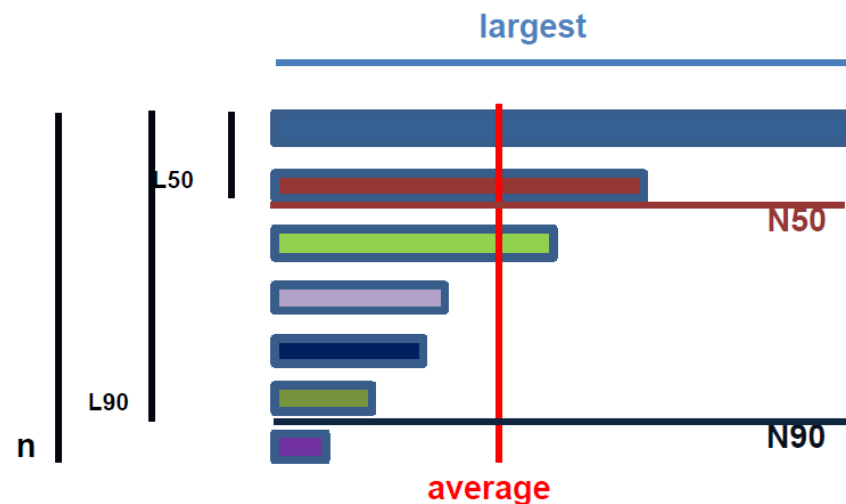
- N50 = length of the shortest contig where 50% of sum is held
- N90 = length of the shortest contig where 90% of sum is held
- L50 = number of contigs which have 50% of the genome
- L90 = number of contigs which have 90% of the genome

e.g., N50为将所有Contigs按长度从大到小排序后，使得累积长度达到总长度的一半的Contig长度

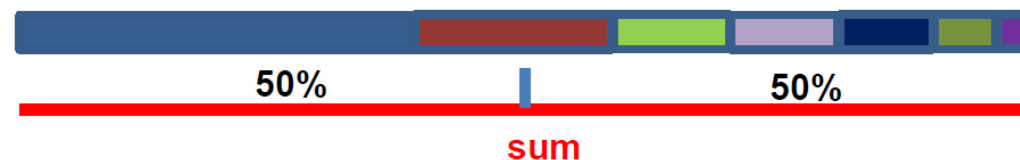


$$190 \rightarrow 50\% \times 190 = 95$$
$$90\% \times 190 = 171$$

- TotalContigLength (70) = 70 < 95
- TotalContigLength (60) = 70 + 60 > 95 → N50:60
- TotalContigLength (30) = 70 + 60 + 30 < 171
- TotalContigLength (20) = 70 + 60 + 30 + 20 > 171 → N90:20



TotalContigLength(t) – total length of all contigs of length at least t



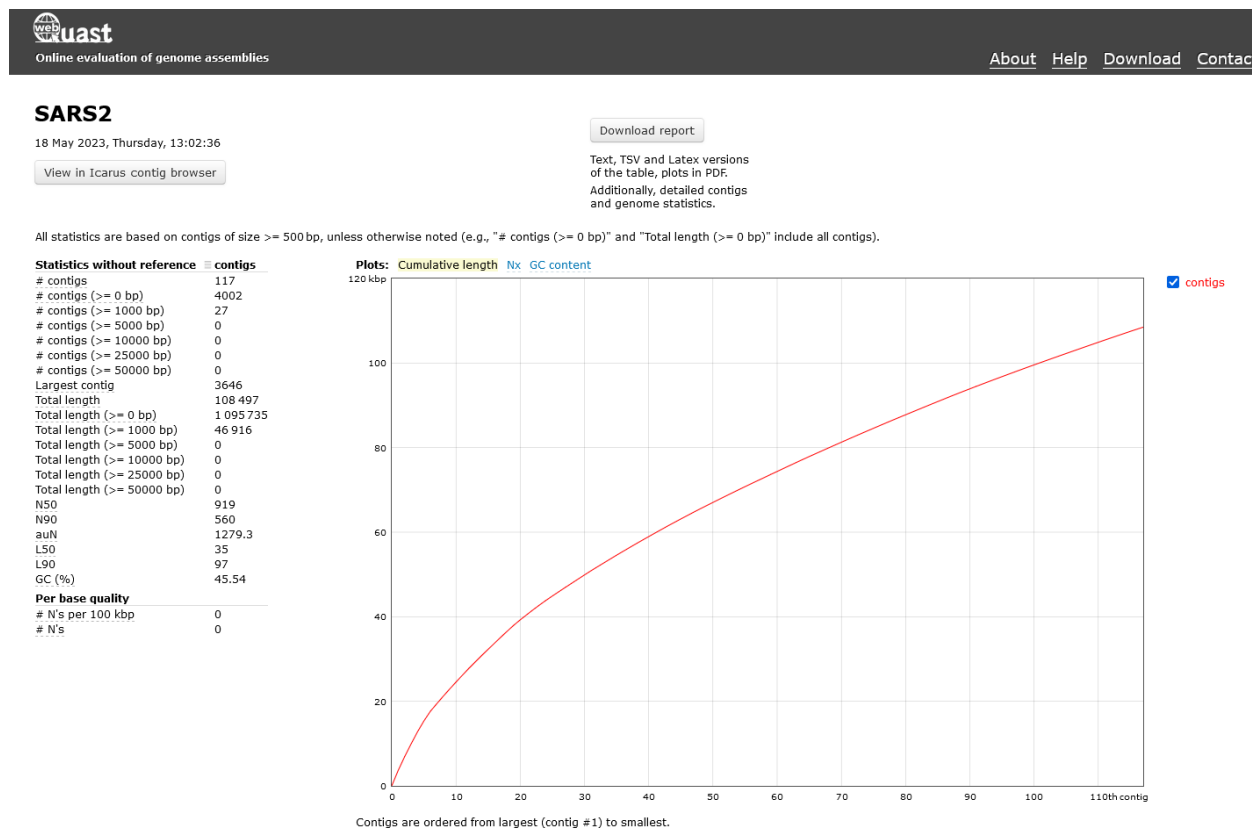
Sum: total length of all contigs

组装质量评估软件

- QUAST (Quality Assessment Tool for Genome Assembly)

- <https://quast.sourceforge.net>

- 根据多种指标，如组装速度、重叠群N50、最大重叠群长度等，评估从头组装软件的效果。



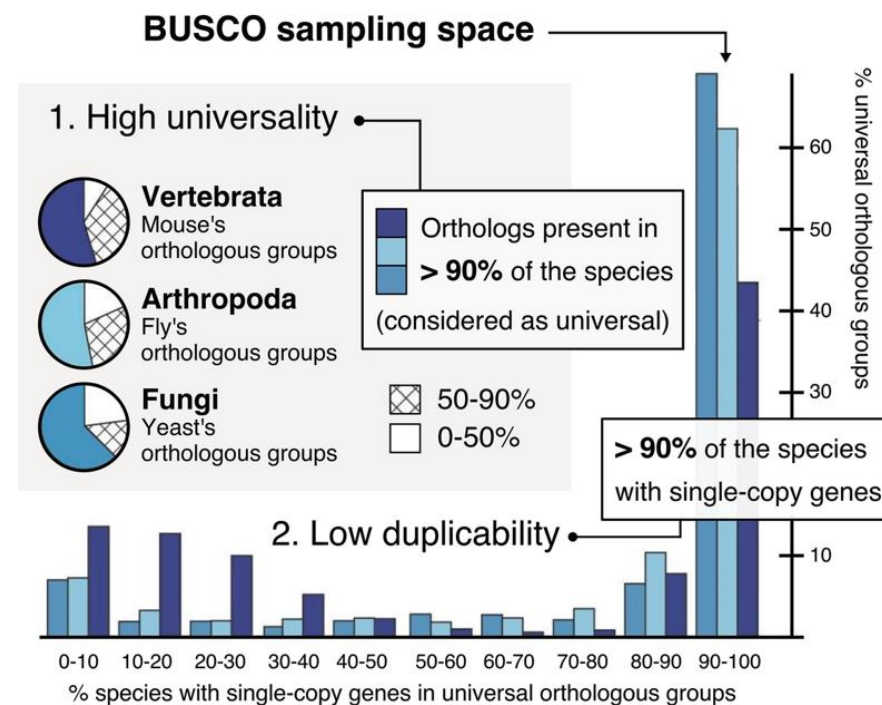
基因组完整性评估

- CEGMA(Core Eukaryotic Genes Mapping Approach)

- CEGMA选取了几个跨度较大的物种（人类、果蝇、线虫、拟南芥、酵母等），并找到这些物种中保守的**458**个基因。
- 理论上，所有物种都有这**458**个基因，所以一个测序完成的基因组如果包含了这些基因，则表明了其基因组序列的完整性。

- BUSCO(Benchmarking Universal Single-Copy Orthologs)

- Based on evolutionarily-informed expectations of gene content of **near-universal single-copy** orthologs, the BUSCO metric is complementary to technical metrics like N50.
- BUSCO对各个物种大类分支都有一个保守基因集合（Bacteria, Eukaryota, Fungi, Plant ...）
- 网址：<https://busco.ezlab.org/>



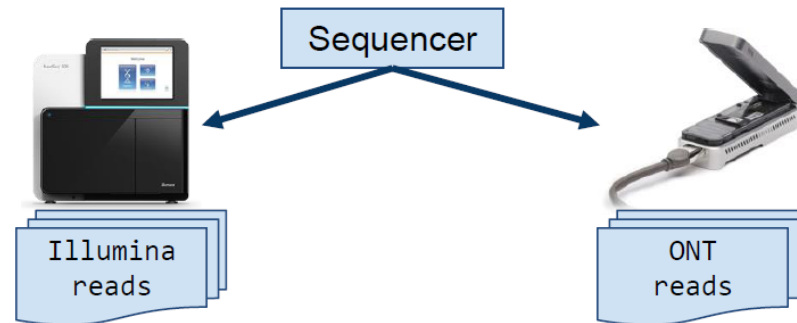


案例：Sequencing of SARS-CoV-2

Sequencing approaches for SARS-CoV-2

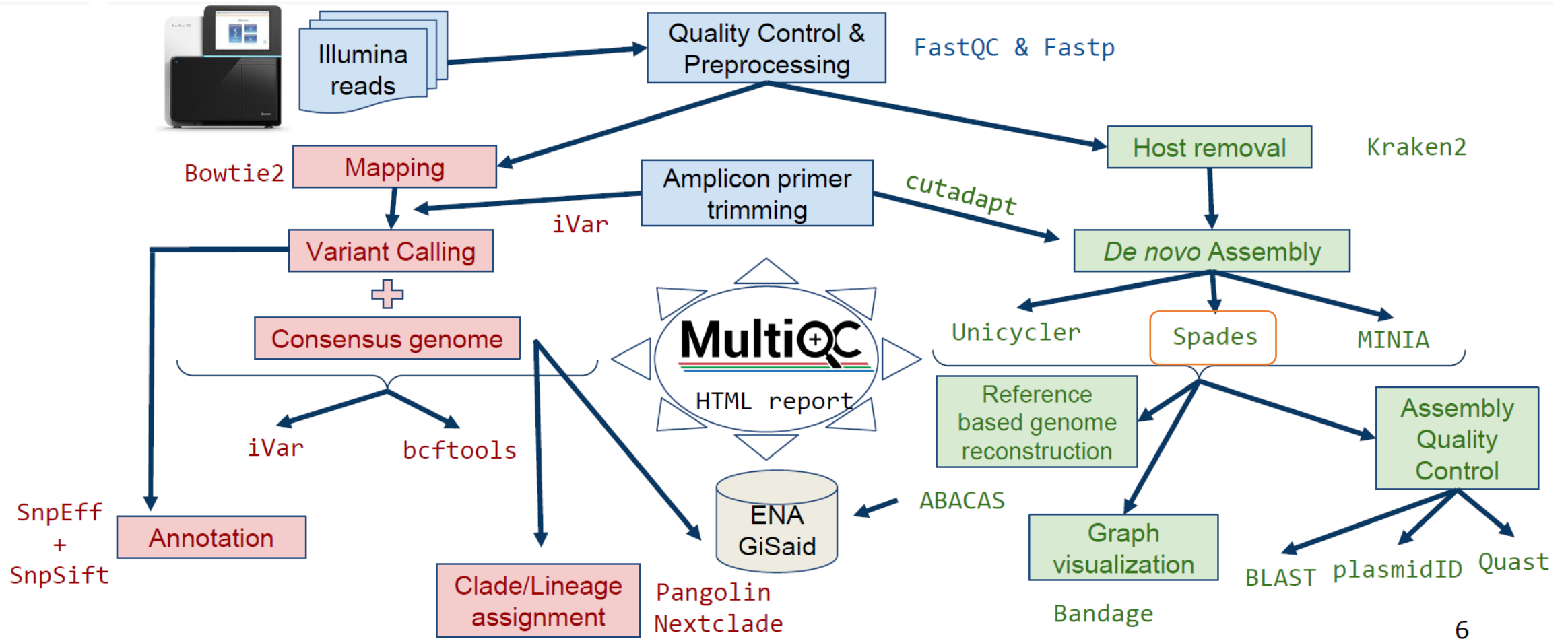
- Current Sequencing Sample Information (<https://galaxyproject.org/projects/covid19/samples/>)

Timeline for 314,941 samples
(Updated March 17, 2022)



[How next-generation sequencing can help identify and track SARS-CoV-2 \(nature.com\)](https://www.nature.com)

Illumina sequencing data analysis



Sequencing of SARS-CoV-2

Article

A new coronavirus associated with human respiratory disease in China

<https://doi.org/10.1038/s41586-020-2008-3>

Received: 7 January 2020

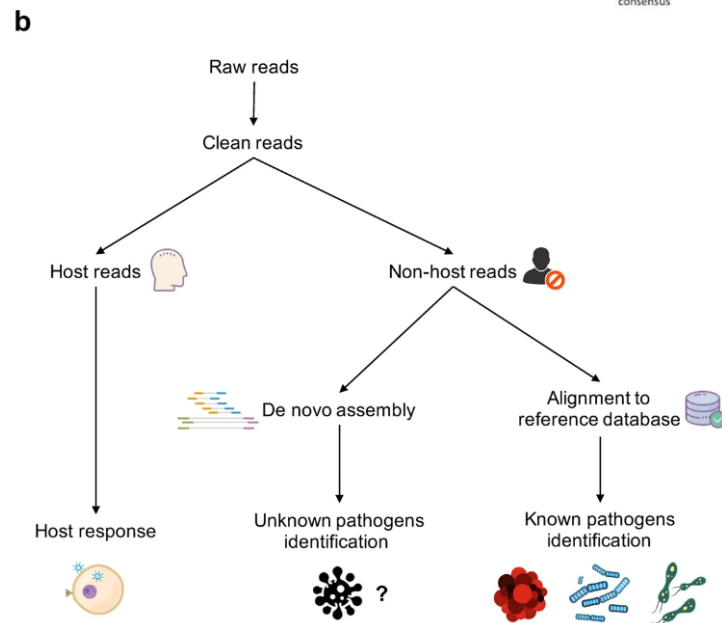
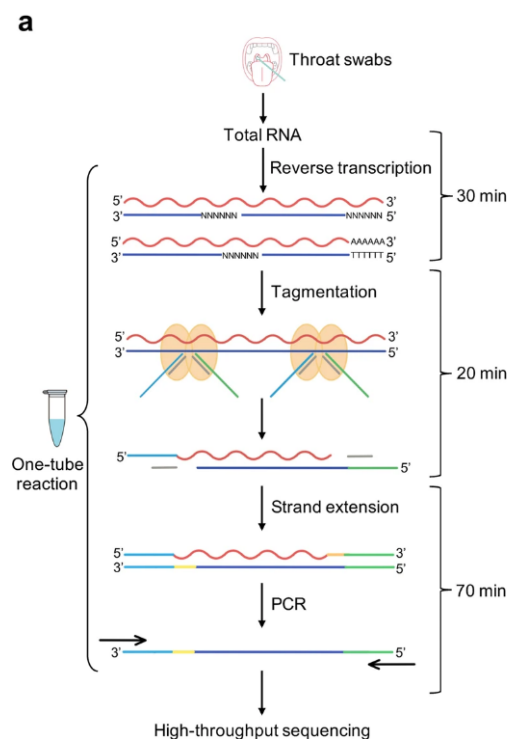
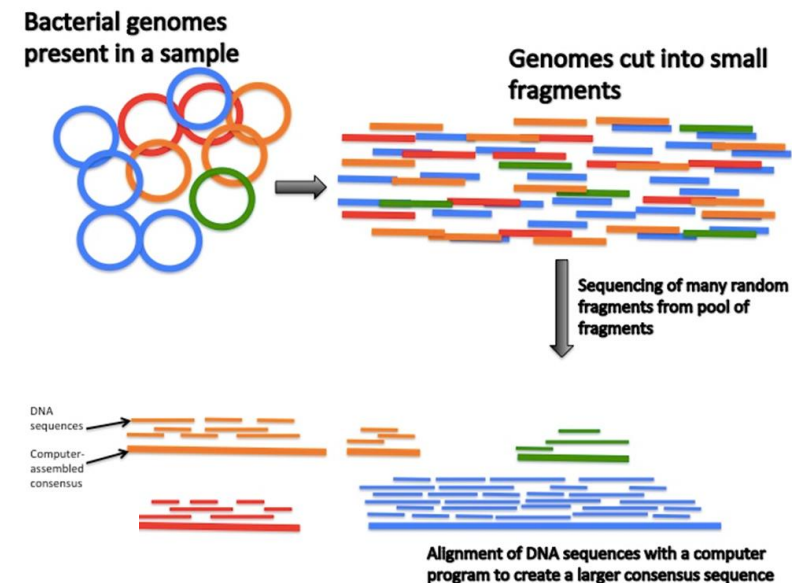
Fan Wu^{1,7}, Su Zhao^{2,7}, Bin Yu^{3,7}, Yan-Mei Chen^{1,7}, Wen Wang^{4,7}, Zhi-Gang Song^{1,7}, Yi Hu^{2,7}, Zhao-Wu Tao², Jun-Hua Tian³, Yuan-Yuan Pei¹, Ming-Li Yuan², Yu-Ling Zhang¹, Fa-Hui Dai¹, Yi Liu¹, Qi-Min Wang¹, Jiao-Jiao Zheng¹, Lin Xu¹, Edward C. Holmes^{1,5} & Yong-Zhen Zhang^{1,4,6}✉

- Total RNA was extracted from the BALF sample of a patient.
 - The term **BALF** is a shorthand for **Bronchoalveolar Lavage Fluid**, which is fluid collected from a patient's lungs.
- It is a mix of the patient's RNA and the viral RNA.
 - this mixture of RNA is called a **metatranscriptome**
- Sequencing data: <https://www.ncbi.nlm.nih.gov/sra/SRR10971381>

Run	# of Spots	# of Bases	Size	Published
SRR10971381	28,282,964	8G	2.6Gb	2020-01-27

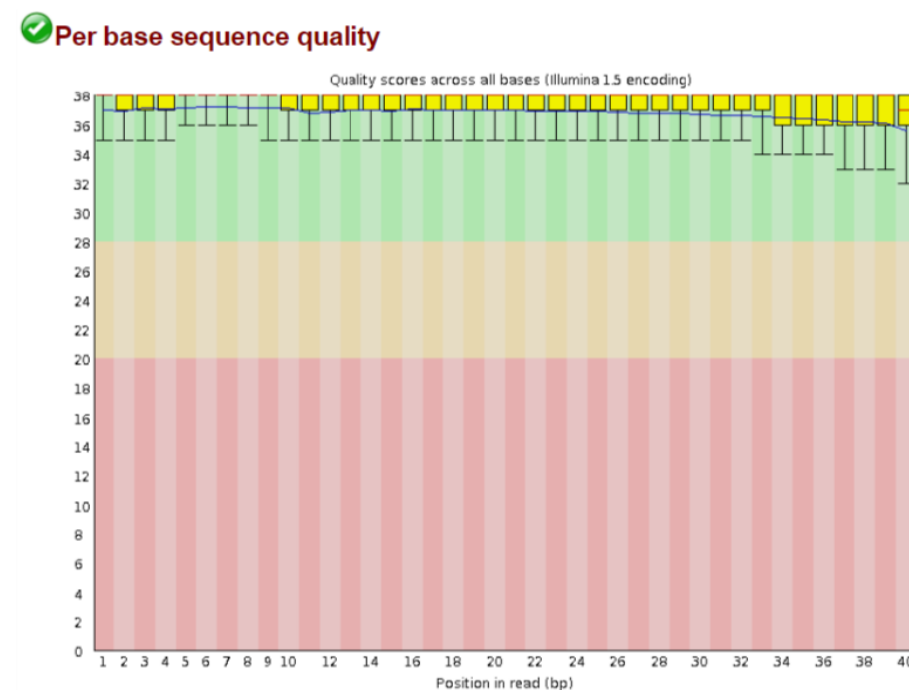
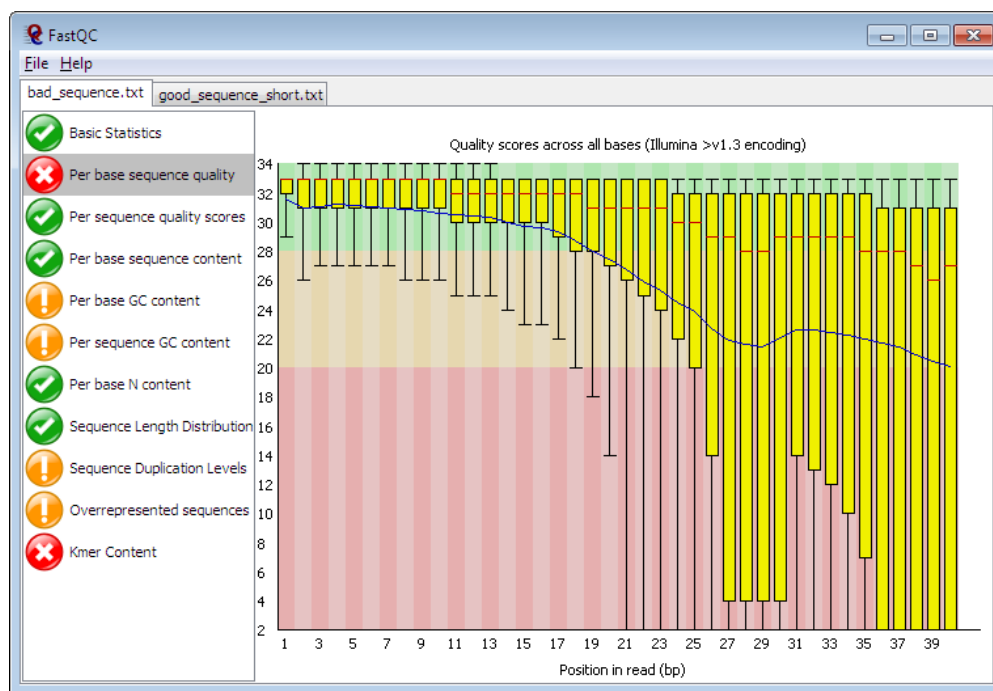
Metagenomic Next-Generation Sequencing

- NGS技术在微生物检测中的应用为宏基因组测序(metagenomic Next-Generation Sequencing, mNGS)。
 - DNA: Metagenome sequencing
 - RNA: Metatranscriptome sequencing
- mNGS是新冠疫情中鉴定出新冠病毒的主要技术。



数据质控-FastQC

- FastQC是目前最常用的NGS数据质量评估软件，可用于统计质量分数、GC含量、测序长度等信息
 - <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>



Overview of the range of quality values across all bases at each position in the FastQ file

数据预处理-Trimomatic

- Trimmomatic 可用于过滤与切除低质量序列、接头序列，不需要的污染物序列等

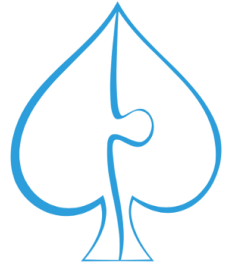
- <http://www.usadellab.org/cms/uploads/supplementary/Trimmomatic/Trimmomatic-0.38.zip>

- 执行命令：

```
$ java -jar Trimmomatic-0.39/trimmomatic-0.39.jar PE SRR10971381_1mini.fq  
SRR10971381_2mini.fq SRR10971381_1_paired.fq SRR10971381_1_unpaired.fq  
SRR10971381_2_paired.fq SRR10971381_2_unpaired.fq ILLUMINACLIP:  
Trimmomatic-0.39/adapters/TruSeq3-PE.fa:2:30:10:2:True LEADING:20  
TRAILING:20 MINLEN:36
```

注意，在运行此java命令的目录下安装Trimmomatic，并存放测序数据！

Assembly with SPAdes



- SPAdes (St. Petersburg genome assembler)是一款非常好用的软件，而且该软件不仅支持Illumina测序数据，而且还可以用于Ion Torrent, PacBio、Sanger, Nanopore测序数据。使用简单，非常适合用于混合组装来改善拼接效果。
 - 安装说明: <http://ablab.github.io/spades/installation.html>
- `$ spades.py -1 SRR10971381_1_paired.fq -2 SRR10971381_2_paired.fq --careful -o sars_illupe`
 - `-o output_dir`: 指定输出的文件夹;
 - `-1/-2`: 单个paired-end library的左端与右端测序reads;
 - `--pe1-1/--pe1-2`: 参数用于质量过滤后还是配对的reads, pe后的数字1为每1个文库的编号, 而后跟着配对reads的编号, 1是左端(forward), 而2是右端(reverse);
 - `--pe1-s`: paired-end library 数据过滤后两端reads不能再配对的single数据, 就在同一个--pe参数后加-s标记;
 - `--careful`: 通过运行MismatchCorrector模块进行基因组上mismatches和short indels的修正, 推荐使用此参数。
- 命令运行结束后, 在输出文件夹sars_illupe中产生许多文件, 其中configs.fasta是最终的组装结果。

Analyzing your assembly

- 用记事本打开contigs.fasta文件，复制第1条序列(NODE_1)做BLAST，结果显示为新冠病毒序列。
- BLAST program: BLASTN
- BLAST parameters:
 - Entrez Query: 1900/12/01:2020/02/01[PDAT] (search results before January 1st, 2020)
 - Organism: viruses

Standard Nucleotide BLAST

blastn blastp blastx tblastn tblastx

BLASTN programs search nucleotide databases using a nucleotide query. [more...](#) [Reset page](#) [Bookmark](#)

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#) Query subrange

From

To

Or, upload file subsampld_contigs.fasta [↩](#)

Job Title

Enter a descriptive title for your BLAST search [↕](#)

Align two or more sequences [↕](#)

Choose Search Set

Database Standard databases (nr etc.): rRNA/ITS databases Genomic + transcript databases Betacoronavirus

Nucleotide collection (nr/nt) [↕](#) [↕](#)

Organism exclude [+](#)

Optional Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown [↕](#)

Exclude Models (XM/XP) Uncultured/environmental sample sequences

Optional

Limit to Sequences from type material

Optional

Entrez Query [↕](#) [YouTube](#) [Create custom database](#)

Optional Enter an Entrez query to limit search [↕](#)

Program Selection

Optimize for Highly similar sequences (megablast)

More dissimilar sequences (discontiguous megablast)

Somewhat similar sequences (blastn)

Choose a BLAST algorithm [↕](#)

[BLAST](#) [↩](#) Search database Nucleotide collection (nr/nt) using Megablast (Optimize for highly similar sequences)

Show results in a new window

BLAST results will be displayed in a new format by default
You can always switch back to the Traditional Results page. [New](#)

QUAST-the Quality Assessment Tool for Genome Assembly

● QUAST本地安装与运行:

- `$wget https://downloads.sourceforge.net/project/quast/quast-5.2.0.tar.gz`
- `$tar -zxvf quast-5.2.0.tar.gz`
- `$/quast-5.2.0/quast.py contigs.fasta -o quast_result`

● QUAST online tool: <http://cab.cc.spbu.ru/quast/>

- "Scaffolds" is not checked,
- "Prokaryotic" is selected,
- "Genome" is set to " SARS-CoV-2"

Quality Assessment

Assemblies File size limit is 100Mb

contigs.fasta 91.0MB [remove](#) ←

Skip contigs shorter than bp

Scaffolds ← (Scaffolds are splitted by fragments of N's ≥ 10 bp)

Find genes

Prokaryotic ← (Process contigs with GeneMarkS, process circular chromosomes)

Eukaryotic (find genes with GeneMark-ES)

Genome ←

[Another genome](#)

Caption

Will appear in the report.

Explore k-mer Length Effects

- Look at the log file generated (spades.log), what k-mer sizes were used for the assembly?
 - Default k-mer sizes were set to [21, 33, 55, 77]

- Try to assembly with a different k-mer length:

```
$k = 127
```

```
$spades.py \
```

```
--threads 2 \
```

```
-k ${k} \
```

```
-o ~/de_novo_illumina/SRR10971381-k${k} \
```

```
-1 ~/data/illumina_pe/SRR10971381_1_paired.fq \
```

```
-2 ~/data/illumina_pe/SRR10971381_2_paired.fq \
```

```
| tee ~/de_novo_illumina/SRR10971381-k${k}.log
```

Hacking COVID-19 — Identifying a Deadly Pathogen

- 参考教学视频:

- <https://www.coursera.org/learn/covid-19-genome-assembly>

The image shows a screenshot of a Coursera course page. At the top, there is a blue header with the Coursera logo. Below the header, the course title 'Hacking COVID-19 — Course 1: Identifying a Deadly Pathogen' is displayed in large white text on a dark blue background. To the right of the title, it says 'Offered By UC San Diego'. Below the title, there is a rating of 4.5 stars based on 30 ratings and 10 reviews. The instructor's name, Mikhail Rayko, is listed with a link to '+4 more instructors'. At the bottom, there is a white button that says 'Enroll for Free' and 'Starts Apr 12', with a link for 'Financial aid available' next to it.


Browse > Health > Health Informatics

Offered By

UC San Diego

Hacking COVID-19 — Course 1: Identifying a Deadly Pathogen

★★★★☆ 4.5 30 ratings · 10 reviews

 Mikhail Rayko [+4 more instructors](#)

Enroll for Free
Starts Apr 12

[Financial aid available](#)

- 根据本课提供的测序数据(SRR10971381)与分析流程，完成SARS-CoV-2基因组测序分析的数据质控(FastQC, Trimmomatic)、基因组组装与评估(SPAdes, QUAST)与病原体鉴定(BLAST)等。



谢谢

THANK YOU

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